



Research Article

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# Optimum dietary level of *Chlorella vulgaris* powder as a feed additive for some blood parameters and digestive enzymatic activities of grey mullet, *Mugil cephalus*



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## Abstract

The present paper aims to shed lights on effect of optimum dietary level of *Chlorella vulgaris* powder (CP) as a feed additive on some blood biochemical parameters (Cholesterol (CHO), triglyceride (TG), total protein (TP), glucose (GLU), glutamic oxaloacetic transaminase (GOT), glutamic pyruvic transaminase (GPT) and lysozyme) and digestive enzymatic activities (amylase, lipase and protease) of the grey mullet (*Mugil cephalus* L). Four experimental diets were supplemented with CP at 0, 5, 10 and 15g kg<sup>-1</sup> diet (CP0, CP, CP10 and CP15). Number of twelve tanks (60L) with three replicates for treatment group (n=10 per tank with average initial weights 14.95±2.01g) and the control group were considered. After 60 days of the feeding trial, fish fed CP5 diet had lower serum CHO and TG levels than did fish fed CP0, CP10 and CP15 diets (P<0.05). There was no significant difference in GLU between fish fed CP5 and CP10 diet (P>0.05). The highest serum total protein, amylase, protease, lipase and lysozyme activities were observed in fish fed CP5. Also, fish fed CP10 and CP15 diets had higher digestive enzymatic activities, serum total protein and lysozyme activities than did fish fed CP0 (P<0.05). These results indicate that dietary supplementation of 5g chlorella powder in the commercial diets could improve the blood chemical responses and the activity of digestive enzyme in grey mullet.

**Keywords:** *Mugil cephalus*; *Chlorella vulgaris*; Additive digestive enzymes; Blood biochemical parameters

## Introduction

Successful rearing of larval fish is the most critical stage in the production cycle for many species. *Mugil cephalus* has a promising market potential in Europe, East and South Asia [1]. It is also an important aquaculture species in Iran. The consumer demand stimulated the development of intensive aquaculture of this species in Asian countries. The problem in rearing larval fish is that of food supply [2]. Therefore a readily available, easily acceptable and highly digestible diet with high nutritional value should be used as larval fish starter diet [3,4].

Plants are natural sources of safer and cheaper chemicals. Plant products have been reported to promote various activities like anti stress, growth promotion, appetite stimulation and immunostimulation in aquaculture practices [5,6].

*Chlorella* sp. as one of the most common microalgae used in aquaculture, have been reported to improve lipid metabolism, digestive enzymatic activities, growth, feed utilization in Korean rockfish, *Sebastes schlegeli* [7], juvenile Japanese flounder, *Paralichthys olivaceus* [8], gibel carp, *Carassius auratus gibelio* [9,10], olive flounder, *Paralichthys olivaceus* [11].

Freshwater algae such as *Chlorella* and *Spirulina* seem to have good potential as protein sources, whereas marine microalgae are the fundamental source of the long-chain polyunsaturated fatty acids that are crucial for human health, as well as that of aquaculture animals. *Chlorella vulgaris* shows some potential as an alternative feed ingredient in aquaculture due to its richness in protein, polysaccharides, vitamins and microelements [9]. Although the algal feeds were not utilized as efficiently as the fishmeal feed, *Chlorella* was well accepted. As example Shi et al. [10] showed that *Chlorella* meal could totally replace dish meal in diet of crucian carp, *C. auratus*.

Until now, no trial has been conducted to study the effect of dietary *Chlorella* powder on some of serum biochemical parameters and digestive enzymatic activities of grey mullet. This study was therefore designed to investigate effect of optimum dietary level of *Chlorella vulgaris* powder (CP) as a feed additive on some blood biochemical parameters (Cholesterol (CHO), triglyceride (TG), total protein (TP), glucose (GLU), glutamic oxaloacetic transaminase (GOT), glutamic pyruvic transaminase

(GPT) and lysozyme) and digestive enzymatic activities (amylase, lipase and protease) of the grey mullet.

## Materials and Methods

### Experimental diets and feeding conditions

*Chlorella vulgaris* powder were purchased from the Yakhteh Nano-chemistry Co in Tehran, Four diets were prepared with

supplementation of *Chlorella* powder (CP) at inclusion level of 0, 5, 10 and 15g kg<sup>-1</sup> diet, analyzed proximate compositions are presented in Table 1. After mixing well with ingredients, oil and 30% distilled water were added and further mixed. The wet dough was pelletized at a particle size of 1mm using a chopper machine (National, Japan). The experimental diets were freeze-dried, sieved into the desired particle size (1mm) and then stored at -4-8 °C until use [12].

**Table 1:** Ingredients (g kg<sup>-1</sup>) and chemical composition (%) of the experimental diets.

Ingredients (g kg <sup>-1</sup> )	Diets			
	CP0	CP5	CP10	CP15
<i>Chlorella vulgaris</i> (g kg <sup>-1</sup> )	0	5	10	15
Fish meal	427	427	427	427
Soybean meal	192.5	192.5	192.5	192.5
Wheat flour	93	93	93	93
Dried yeast	37.5	37.5	37.5	37.5
Fish oil	55	55	55	55
Soy oil	27.5	27.5	27.5	27.5
Choline chloride	2	2	2	2
Bi calcium phosphate	3.7	3.7	3.7	3.7
Lecithin	28.15	28.15	28.15	28.15
Premix <sup>a</sup>	9.4	9.4	9.4	9.4
<b>Proximate composition (%)</b>				
Crude protein	51.6	51	50.6	51.6
Crude lipid	11.9	11	11.4	11.2
Crude ash	12.1	12	11.8	12.6
Dry matter	92.2	92.1	92	92

<sup>a</sup>Premix (mg kg<sup>-1</sup>) KI, 250; MnSO<sub>4</sub>·H<sub>2</sub>O, 2800; ZnSO<sub>4</sub>·H<sub>2</sub>O, 2350; vitamin K, 225; biotin 3500, (2%) niacin, 4850; calcium pantothenate, 11,000; folic acid, 2000; vitamin B<sub>1</sub>, 1500; vitamin B<sub>2</sub>, 2000; vitamin B<sub>6</sub>, 2000; and vitamin C, 50,000.

The feeding experiment was conducted at Fisheries Research Center, Chabahar, Iran. one hundred- twenty grey mullet with an initial mean weight of 14.95g were randomly distributed into twelve tanks (60L) at a stocking density of 10 fish/tank (triplicates per treatment) and fed to satiation by hand twice (09:00 and 17:00) a day for 60 days and feed intake was recorded on daily basis. Dissolved oxygen concentration, ammonia nitrogen concentration and pH were measured about 7.01±0.87mg L<sup>-1</sup>, 0.11±0.04mg L<sup>-1</sup> and 7.8±0.4 respectively. The photoperiod was regulated as a 12:12h (dark/light) cycle.

### Biochemical analysis

At the end of experiment, number of nine fish from each treatment were anesthetized (with clove oil at 5mg L<sup>-1</sup>) and blood samples were taken after excising caudal peduncle and were transferred to un-heparinized sterile tubes 1-1.5mL for the serum biochemical tests proposes [2,13].

Serum glucose concentration was measured according to method described By Trinder [14]. Serum total protein was determined via the method described by Wootton [15]. Triglyceride and cholesterol level determination was carried out

according to the method described by Sankar [16]. Biochemical estimation of blood glucose, protein, cholesterol and triglyceride were determined by means of standard analyses kits (Pars Azmon, Iran) using automatic analyzer (Furuno, CA-270, Japan).

The turbidimetric assay for lysozyme was carried out with minor modification [17]. In brief, substrate for the assay of lysozyme was performed using 0.03% of lyophilized cells of *Micrococcus lysodeikticus* (Sigma, ATCC No. 4698) in 0.05mM sodium phosphate buffer (pH 6.2). Twenty-five microliters of fish plasma were added to 175µl bacterial suspensions in duplicate wells of a microtitre plate. The mixture was incubated at room temperature and an absorbance at 600nm was measured after 15s using an ELISA plate reader (Argus, Perkin Elmer, France). A unit of lysozyme activity was defined as a plasma-decrease amount of lysozyme in absorbance of 0.001 mL<sup>-1</sup> min<sup>-1</sup>.

### Digestive enzyme activity

For preparation of enzyme extracts, three fish from each tank were randomly selected and sacrificed. The digestive tracts were carefully dissected out, thoroughly washed with sterile distilled water, weighed and separately homogenized with cooled buffer

phosphate (0.65%, pH 7, 1: 10w/v). The supernatant, extracted by centrifugation (3000g for 20min at 4 °C) (Centrifuge EBA21, Hettich, Germany), was used for enzyme assays. Amylase activity was assessed by 3, 5- dinitrosalicylic acid (DNS) method (King, 1965). 0.1ml tissue homogenate, 2ml phosphate buffer (0.1M, pH 7) and 0.1ml of 1% (w/v) starch solution was mixed and incubated at 30 °C for 35min. Then adding 2ml DNS reagent stopped the reaction. After 5min in boiling water, the reaction mixture was cooled, diluted with distilled water and recorded the absorbance at 540nm. Protease activity was measured by the casein digestion method of king (1965). 0.1ml tissue homogenated, 0.05M tris phosphate buffer (PH 7.8), 0.01N NaOH and 2.5ml of 1% (w/v) consisted the reaction mixture. The mixture was incubated at 30 °C for 10min and stopped by 2.5ml, 10% trichloroacetic acid (TCA) and filtered. The reagent blank consisted just tissue homogenate before stopping the reaction and without incubation. The absorbance was recorded at 320nm. Unit amylase activity was calculated as the weight (mg) of maltose liberated for a duration of 10min at 30 °C. Unit protease activity was expressed as the amount of tyrosine liberated in 15min under the assay conditions. Lipase activity was assessed by King method. Olive oil emulsion, phosphate buffer (pH 7.8, 0.1M), tissue homogenate and distilled water consisted the reaction mixture. The reaction mixture was incubated at 30 °C for 24h and added two drops phenolphthalein indicator and 95% alcohol for titration against 0.05N NaOH until the appearance of permanent pink color. Unit lipase activity expressed as the amount of 0.025N NaoH required to neutralize

the fatty acids liberated during 18h of incubation at pH 6.9 and temperature 30 °C. Digestive enzymes were calculated as enzyme unit per gram tissue.

### Statistical analysis

All measurements were repeated twice. Data were evaluated using one- way analysis of variance (ANOVA). Groups were considered to be significantly different if P<0.05. When a significant F value was obtained for ANOVA the differences between all groups were tested by using Duncan multiple comparisons test. All statistics were performed using SPSS for windows versions 16. Data are reported as means±standard Error.

### Results

#### Effect of Chlorella powder (CP) on the blood biochemical parameters of grey mullet

The blood parameters of fish fed with the experimental diets were shown in Table 2. Analysis of the data in Table 2 suggested that the dietary chlorella mainly affected the parameters involved in protein/ lipid metabolism and immunity of grey mullet. The serum total protein content and lysozyme activity were significantly (p<0.05) increased for fish fed CP diet over the control. The highest total protein content was recorded in those fish fed CP5 diet. There was significant (p<0.05) decline in cholesterol, glucose and triglycerides levels in those fish fed CP diet over the control (Table 1). The lowest cholesterol, glucose and triglycerides levels of serum were observed in fish fed CP5.

**Table 2:** Serum biochemical parameters of *M.cephalus* fed CP diet at different levels for 60 days.

Parameter	CP diet (g kg <sup>-1</sup> feed)			
	0	5	10	15
Total protein (g dl-1)	4.42±0.43 <sup>d</sup>	6.58± 0.33 <sup>a</sup>	5.45±0.21 <sup>b</sup>	4.97±0.18 <sup>c</sup>
Glucose (mg dl-1)	55±1.52 <sup>a</sup>	31±11.52 <sup>c</sup>	35.33±10.45 <sup>bc</sup>	37±11.33 <sup>b</sup>
Triglycerides (mg dl-1)	227.33±11.76 <sup>a</sup>	172±10.05 <sup>d</sup>	191±12.64 <sup>c</sup>	209.28±14.75 <sup>b</sup>
Cholesterol (mg dl-1)	104.79±10.76 <sup>a</sup>	75.33±13.7 <sup>c</sup>	95±11.73 <sup>b</sup>	100±11.15 <sup>b</sup>
Lysozyme	149±15.53 <sup>c</sup>	275.07±18.45 <sup>a</sup>	199.67±10.88 <sup>b</sup>	185±12.88 <sup>b</sup>

CP diet, *Chlorella* powder diet. Values (mean±SE of three replication). In each row not sharing a common superscript are significantly different (P<0.05).

#### Effect of Chlorella powder (CP) on the digestive enzymatic activities of grey mullet

The activity of three digestive enzymes including amylase, lipase and protease in intestine were examined, and the results were summarized in Table 3. These three enzymes were

significantly increased for fish fed CP diet compared with control. The highest the digestive enzymatic activities were recorded in those fish fed CP5 diet. However there was no significant changes in the activity of these three enzymes among fish fed CP10 diet and CP15 diet (P>0.05).

**Table 3:** Digestive enzyme activities of *M. cephalus* fed CP diet at different levels for 60 days.

Specific activity of enzyme (unit mg <sup>-1</sup> protein)	CP diet (g kg <sup>-1</sup> feed)			
	0	5	10	15
Amylase	26.3±8.92 <sup>c</sup>	45.66±11.45 <sup>a</sup>	34.33±9.32 <sup>b</sup>	36±12.08 <sup>b</sup>
Protease	356.67±23.07 <sup>c</sup>	520±11.73 <sup>a</sup>	450±15 <sup>b</sup>	432±14.93 <sup>b</sup>
Lipase	211±10.21 <sup>b</sup>	514±11.58 <sup>a</sup>	406.67±17.67 <sup>b</sup>	378.67±16.90 <sup>b</sup>

Cp diet, *Chlorella* powder diet. Values (mean±SE of three replication). In each row not sharing a common superscript are significantly different (P<0.05).

## Discussion

In the present study, the different contents of *Chlorella* powder were added in the basal diet of grey mullet and the effects of *Chlorella* on the blood biochemical parameters and digestive enzyme were detected.

5-15g *Chlorella* powder in diets could significantly improve the blood parameters of grey mullet. In this study, significantly lower serum total cholesterol concentrations were detected in all levels of CP in comparison to those fed the control diet. In agreement with our results, Rahimnejad & Lee [11] reported the significant reduction of serum cholesterol level in olive flounder, *Paralichthys olivaceus* fed diets containing 5-15% *C. vulgaris*. Also Xu et al. [9] found the significant decrease of serum cholesterol level in gibel carp fed diets supplemented with 1.6-2% *Chlorella* powder. Similar tendency of decreasing blood cholesterol level also been achieved in olive flounder fed diets containing 2-4% *C. ellipsoidea* [8], suggesting that *Chlorella* supplementation could activate hormonal regulation of lipid metabolism [9]. In this study, significantly higher serum total protein concentrations and lysozyme activity were detected in all levels of CP in comparison to those fed the control diet. Similar tendency of increasing the serum total protein and lysozyme activity also been achieved in olive flounder fed diets containing 2-4% *C. ellipsoidea* and in gibel carp fed 1.6-2% *Chlorella* powder. Protein and lysozyme play important roles in the immune system [9,18]. Our findings suggested that the dietary adding *Chlorella* could increase the immune response of grey mullet. Lysozyme is also an important defense molecule of the innate immune system, which is important in mediating protection against microbial invasion [18]. Increasing lysozyme and total protein by dietary *Chlorella* suggested that the *Chlorella* might contain some bioactive substances involving in the regulating of fish immune response.

In this study, significantly lower triglyceride and glucose levels were detected in all levels of CP in comparison to those fed the control diet, suggesting that the *Chlorella* might involve in the metabolism of carbohydrate. Conversely, Xu et al. [9] showed that *Chlorella* powder could decrease the level of blood cholesterol not the glucose of gibel carp. The same case also found by Güroy et al. [19], suggesting that its effect also variably depend on dietary *Chlorella* species and its concentrations [8].

The results of the present study indicated that using the different levels *C. vulgaris* powder play a positive role on the activity of digestive enzymes. Analysis of digestive enzyme activity is an easy and reliable methodology that can be used as an indicator of digestive processes and nutritional condition of fish [20]. *M. cephalus* is a stomach- less fish. Digestion takes place in the intestine, in which various intestinal enzymes are involved in digestive and absorptive processes, such as amylase, protease, lipase [21]. Similar positive observations were reported by Xu et al. [9] with increasing the digestive enzyme in the hepatopancreas and intestine in gibel carp, *Carassius auratus gibelio* fed with *Chlorella*, suggesting the *Chlorella* could enhance

the diet utilization rate by increasing the activity of digestive enzyme. Also, Shi et al. [10] showed that dietary *Chlorella* meal substitution could significantly increase the activities of amylase but not activities of trypsin and lipase in intestine of crucian carp, *Carassius auratus*. Increased amylase activities may improve the utilization of carbohydrates in diets. Vizcaino et al. [22] reported that algae inclusion could significantly increase the activities of trypsin but not activities of trypsin of sea bream, *Sparus aurata*. In this study, the highest digestive enzyme activities were observed in fish fed with CP5, while the activity of these three enzyme in gibel carp fed with 2% *Chlorella* group were the higher than those of other groups. Suggesting the optimum level of dietary microalga on digestive enzymatic activities was variable even in conspecific [9]. Information about suitable concentration of dietary microalga that beneficially affects fish digestive enzymes might be required for application in aquafeed.

## Conclusion

In conclusion, the present study revealed that dietary supplementation of 5g *C. vulgaris* powder in each kg diet, as a feed additive, improved biochemical parameters and digestive enzymatic activities in grey mullet. Therefore, this study suggests that optimum dietary supplementation level of *Chlorella* powder could be approximately 5g kg<sup>-1</sup> of diet for the positive effects on blood biochemical parameters and digestive enzymatic activities in grey mullet.

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