Effect of *Origanum vulgare* Extract on Immune Responses and Haematological Parameters of Rainbow Trout (*Oncorhynchus mykiss*)

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Submission: December 13, 2017; Published: February 15, 2018

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Abstract

In this study, *Origanum vulgare* extract was used to evaluate its effects on immune responses and haematological parameters of rainbow trout (*Oncorhynchus mykiss*). Six hundred (600) average mean weight 13±0.05g rainbow trout (*Oncorhynchus mykiss*) were randomly allocated into two groups including placebo-treated group (control), and *Origanum vulgare* extract-treated group, each of three replicates. The fishes were hand-fed once a day with diet medicated placebo or *Origanum vulgare* extract (OE) at a rate of 1% of feed weight in the first feeding for 8 weeks. At the end of every two weeks 24hrs after feeding, fish were bled from caudal vein and blood samples were analyzed for some of haematological and immunological parameters. The results showed that serum total protein, albumin and globulin, respiratory burst activity, phagocytic activity and serum lysozyme activity vary among the two treatment groups which were found to be higher in OE-treated group (P<0.05). It was concluded that supplementation of OE at a rate of 1% registered higher immunological responses. Therefore, dietary inclusion of OE could improve non-specific immune responses in rainbow trout. Future studies to determine optimal herb mixtures and dietary levels should be conducted.

Keywords: Herbal immunostimulant; Iranian medicinal plants; *Origanum vulgare*; Fish

Introduction

Major targets in the aquaculture industry are to maintain fish health as well as to improve fish performance. The use of plant extracts in practical diets for fish is a very topical concept in aquaculture industry. One of the most important aspects in rainbow trout farming is the nutrition factor. It can influence the performance as well as the health status of the cultured fish. *Origanum vulgare* is a member of the Labiatae family of plants. It is an aromatic plant with a wide distribution throughout the Mediterranean area and Asia [1]. The essential oil obtained from *O. vulgare* subsp. *hirtum* plant by a steam distillation process comprises more than 20 ingredients, most of which are phenolic antioxidants [2]. Major components are carvacrol and thymol that constitute about 78 to 82% of the total oil [3]. It has been suggested that the essential oil derived from oregano possess *in vitro* antimicrobial [4,5] antifungal [6], insecticidal [7] and antioxidant [8] properties. These properties are mainly attributed to carvacrol and thymol. The activity of other constituents such as the two monoterpenic hydrocarbons, γ-terpinene and p-cymene, that often constitute about 5 and 7% of the total oil, respectively [3] is uncertain.

Materials and Methods

Preparation of *Origanum vulgare* extract

The plant of *Origanum vulgare* was procured from local store and plant species was identified and confirmed by a botanist in Institute of Medicinal Plants. The dried air parts were collected and washed in sterile distilled water. The samples were separately shade-dried for 10 day till weight constancy was achieved. Then, the samples were powdered in an electric blender. The extract was prepared with the standard method of percolation. To do this, chopped dried air parts of plant in 80% ethanol were percolated for 72 hours. Then, the slurry was filtered with Whatman No. 1 filter paper and centrifuged for 5min at 5000rpm. The filtrate obtained from ethanol using a rotary device, the excess solvent was separated from the extract. These crude extract was stored at 4 °C until use.
Supplementation of the normal diet with dried *Origanum vulgare* extract

The formulated fish feed was prepared using the normal fish diet (50% crude protein, 18% crude lipid, 1.9% fiber, 1.3% total phosphorus, 8.3% ashes, and 14.8% nitrogen free extract) with dried *Origanum vulgare* extract or placebo at a ratio 1% of weight food and mixing part by part in a drum mixer. Sufficient water along with the oil ingredients were then added to make a paste of each diet. After it was pelleted and allowed to cool dry. The pellets were air dried and stored in air tight containers until fed.

Fish and experimental conditions

600 rainbow trout weighing 13±0.05 were used. All experiments were carried out in 1,000 liter round concrete ponds with a continuous water flow of 2.5 liter per second. The fish were kept at an ambient, including uncontrolled water temperature of 15±1 °C, dissolved oxygen of 7.2±0.2 mg l⁻¹ and pH 8±0.3. After 2 weeks adaptation, fish were randomly allotted in 6 concrete ponds each containing 100 fish. Each group was hand-fed once a day with diet medicated 1% of *Origanum vulgare* extract, or placebo (70% lactose, 10 % starch and 20 % talc) prepared in the laboratory and three times with normal diet at a rate of 2% of body weight for 10 weeks.

Bleeding and serum collection

During bleeding, fish were rapidly netted, tranquillized with 50mg/l of tricaine methane sulfonate (MS₂₂₂ Sigma chemical Co. St. Louis, MO, USA). Fish were bled from caudal vein using 1ml insulin syringe fitted with 24 gauge needle. To minimize the stress to fish, 1ml of blood was drawn and the whole bleeding procedure was completed within 1min. A total number of 15 blood samples were collected from 15 fish in each group (5 samples from each replicate) at the end of every 2 weeks, 24h after final feeding period. The blood pooling of 5 fish from each replicate divided into 2 haves. Half collected in serological tubes containing a pinch of lithium heparin powder, shaken gently and kept at 4 °C to test hematological parameters. Other half collected in tubes without of anticoagulant and allowed to clot at 4 °C for 2hrs to test serological parameters. The clot was the spun down at 2000rpm for 10min to separate the serum. The serum collected by micropipette and was stored in sterile Eppendorf tubes at -20 °C until used for assay.

Hematological assay

Blood sample was analyzed with routine methods adopted in fish hematology [9]. The total red blood cell counts (RBC ×10⁶/μl) were determined in a 1:200 dilution of the blood sample in Hayem’s solution and total white blood cell counts (WBC ×10³/μl) in a 1:20 dilution of the blood sample with a Neubauer hemocytometer. The hematocrit (Hct) and leucocrit percentages were determined in duplicate by using micro hematocrit-heparinized capillary tubes of 75μl volume and a micro hematocrit centrifuge at 15000rpm for 5min [10]. The percentages of erythrocyte (hematocrit) and leucocyte (leucocrit) volumes were calculated by overlaying the tubes on a sliding scale hematocrit reader.

The hemoglobin (Hbg/dl) concentrations were determined by the cyanomethaemoglobin method [11] using a haemoglobin reagent set (Ziest Chem Diagnostics). The all the values of red blood cell indices, the mean values of cell haemoglobin (MCH pg), cell hemoglobin concentration (MCHC %), and cell hemoglobin volume (MCV fl) were calculated according to Wintrobe formulae [12]. The differential leucocytes count was carried out using blood smears stained with Wright-Giemsa. The percentage composition of leucocytes was determined based on their identification characters listed by Ivanava [13].

Biochemical assay

Serum total protein content was estimated photo metrically by citrate buffer and bromocresol green (BCG) dye binding method [14] using the kit (total protein and albumin kit, Pars Azmun Company, Iran). Albumin was determined BCG binding method. The absorbance of standard and test were measured against blank in a spectrophotometer at 546nm. Globulin level was calculated by subtracting albumin values from total serum protein. Albumin/globulin (A/G) ratio was calculated by diving albumin values by globulin values.

Immunological assay

Separation of leukocytes from the blood

Leucocytes for assay were separated from each blood sample by density-gradient centrifugation. One milliliter of histopaque 1.119 (Sigma) containing 100μl of bactohemagglutination buffer, pH 7.3 (Difco, USA) was dispensed into siliconised tubes. 1ml of a mixture of 1.077 density histopaque and hemagglutination buffer and 1ml of blood was carefully layered on the top. The sample preparations were centrifuged at 2500rpm for 15min at 4 °C. After centrifugation, plasma was collected and stored at -80 °C for future analysis; separated leucocytes were gently removed and dispensed into siliconised tubes, containing phenol red free Hanks Balanced Salt Solution (HBSS, Sigma). Cells were then washed twice in HBSS and adjusted to 2×10⁶ viable cells/ml.

Respiratory burst activity

Respiratory burst activity of isolated leucocytes was quantified by reduction of ferricytochrome C [15]. Briefly, 100μl of leucocyte suspension and an equal volume of cytochrome C (2mg/l in phenol red free HBSS) containing phorbol 12-myristate 13-acetate (PMA, Sigma) at 1μg/ml were placed in triplicate in micro titer plates. In order to test specificity, another 100μl of leucocyte suspensions and solutions of cytochrome c containing PMA and superoxide dismutase (SOD, Sigma) at 300U/ml.
were prepared in triplicate in micro titer plates. Samples were then mixed and incubated at room temperature for 15 min. Extinctions were measured at 550 nm against a cytochrome C blank in a multiscan spectrophotometer. Readings were converted to nmoles O₂ by subtracting the O.D. of the PMA/SOD treated supernatant from that treated with PMA given alone for each sample, and converting O.D. to n moles O₂ by multiplying by 15.87. Final results were expressed as nano moles O₂ produced per 10⁶ blood leukocytes.

**Phagocytosis assay**

Phagocytosis activity of blood leukocytes was determined spectrophotometrically according to Seeley et al. [16]. This assay involves the measurement of Congo red-stained yeast cells which have been phagocytised by cells. To perform the assay, 250 µl of the yeast solution was mixed with 500 µl of the Congo red-stained and autoclaved yeast cell suspension (providing a yeast cell: leukocyte ratio of 40:1). The mixtures were incubated at room temperature for 60 min. Following incubation, 1 ml ice-cold HBSS was added and 1 ml of histopaque (1.077) was injected into the bottom of each sample tube. The samples were centrifuged at 2500 rpm for 5 min to separate leukocytes from free yeast cells. Leukocytes were harvested and washed two times in HBSS. The cells then were resuspended in 1 ml trypsin-EDTA solution (5.0 g/l trypsin and 2.0 g/l EDTA, Sigma) and incubated at 37 °C overnight. The absorbance of the samples was measured at 510 nm using trypsin-EDTA as a blank.

**Serum lysozyme assay**

In this study, an assay based on the lysis of *Micrococcus lysodeikticus* was used to determine the lysozyme activity.

**Results**

Table 1: The haematological parameters, WBC, RBC, Hct, Hb, MCH, MCV, MCHC, neutrophil, monocyte, and lymphocyte of rainbow trout fed with 1% placebo (PL) or *Origanum vulgare* extract (OE) in feed for 8 weeks.

<table>
<thead>
<tr>
<th>Week</th>
<th>Groups</th>
<th>WBC (10³/mL)</th>
<th>RBC (10⁶/mL)</th>
<th>Hct(%)</th>
<th>Hb(g/dL)</th>
<th>MCH(pg)</th>
<th>MCV(fL)</th>
<th>MCHC(%)</th>
<th>Neut(%)</th>
<th>Mon(%)</th>
<th>Lymph(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2</td>
<td>PL</td>
<td>3.90±0.7</td>
<td>0.98±0.6</td>
<td>27.33±3.3</td>
<td>9.07±0.6</td>
<td>92.55±27</td>
<td>278.5±16</td>
<td>3.31±0.34</td>
<td>7.66±0.1</td>
<td>3.00±0.3</td>
<td>89.0±1.2</td>
</tr>
<tr>
<td></td>
<td>OE</td>
<td>3.92±0.2</td>
<td>1.08±0.8</td>
<td>27.77±2.2</td>
<td>9.32±0.9</td>
<td>86.29±4.6</td>
<td>257.1±11</td>
<td>3.35±0.5</td>
<td>8.33±0.1</td>
<td>2.66±0.1</td>
<td>90.0±1.0</td>
</tr>
<tr>
<td>4</td>
<td>PL</td>
<td>4.00±0.1</td>
<td>1.69±0.5</td>
<td>40.50±4.1</td>
<td>13.50±0.9</td>
<td>79.88±31</td>
<td>239.6±13</td>
<td>3.33±0.3</td>
<td>7.66±0.5</td>
<td>3.33±0.2</td>
<td>89.0±2.0</td>
</tr>
<tr>
<td></td>
<td>OE</td>
<td>4.06±0.6</td>
<td>1.71±0.4</td>
<td>40.50±5.5</td>
<td>13.56±1.5</td>
<td>79.29±29</td>
<td>236.8±9</td>
<td>3.33±0.4</td>
<td>7.66±0.4</td>
<td>2.00±0.5</td>
<td>90.3±0.7</td>
</tr>
<tr>
<td>6</td>
<td>PL</td>
<td>4.20±0.5</td>
<td>1.69±0.3</td>
<td>36.00±3.7</td>
<td>12.00±1.5</td>
<td>71.00±3.0</td>
<td>213.0±6</td>
<td>3.33±0.3</td>
<td>7.66±0.3</td>
<td>3.33±0.3</td>
<td>89.0±2.1</td>
</tr>
<tr>
<td></td>
<td>OE</td>
<td>4.10±0.2</td>
<td>1.76±0.6</td>
<td>37.00±2.0</td>
<td>12.06±0.8</td>
<td>68.52±19</td>
<td>210.2±6</td>
<td>3.25±0.2</td>
<td>8.33±0.2</td>
<td>3.00±0.2</td>
<td>88.6±0.9</td>
</tr>
<tr>
<td>8</td>
<td>PL</td>
<td>4.95±0.1</td>
<td>1.53±0.3</td>
<td>33.00±3.5</td>
<td>10.98±1.8</td>
<td>71.76±39</td>
<td>215.6±8</td>
<td>3.32±0.2</td>
<td>8.00±1.0</td>
<td>3.33±0.2</td>
<td>89.6±1.2</td>
</tr>
<tr>
<td></td>
<td>OE</td>
<td>5.31±0.5</td>
<td>1.46±0.6</td>
<td>36.66±3.0</td>
<td>12.33±1.3</td>
<td>84.45±41</td>
<td>251.0±14</td>
<td>3.33±0.3</td>
<td>7.66±0.5</td>
<td>2.00±0.6</td>
<td>90.0±0.6</td>
</tr>
</tbody>
</table>

Data are expressed as mean±SE (n=15). No significant differences were observed in the *Origanum vulgare* treated groups relative to the placebo group at the end of the identical every two weeks after feeding (P>0.05). Neut: neutrophil; Mon: Monocyte; Lymph: Lymphocyte.

**Biochemical analysis**

*Origanum vulgare* extract had significant (p<0.05) effect in increase of total protein (TP), albumin (AL), and globulin (GL), at the end of the identical every two weeks after feeding in compared to placebo group (Table 2). The maximum level of total protein was recorded on week 2 of exposure duration. Similarly, albumin and globulin contents were significantly higher in Aloe vera group as compared to placebo group. However, albumin/
globulin ratio was not exhibited significant differences in compared to placebo group at the end of the identical every two weeks after feeding in compared to placebo group (p>0.05; Table 2).

Table 2: Changes in the serum total protein, albumin, globulin and albumin/globulin ratio of rainbow trout after feeding with 1% placebo, or *Origanum vulgare* extract for 8 weeks.

<table>
<thead>
<tr>
<th>Week</th>
<th>Groups</th>
<th>Total protein (g/dL)</th>
<th>Albumin (g/dL)</th>
<th>Globulin (g/dL)</th>
<th>Albumin/globulin ratio (g/dL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2</td>
<td>Placebo</td>
<td>4.10±0.01</td>
<td>1.80±0.08</td>
<td>2.20±0.08</td>
<td>0.81±0.01</td>
</tr>
<tr>
<td></td>
<td><em>Origanum extract</em></td>
<td>4.23±0.01</td>
<td>1.90±0.07</td>
<td>2.33±0.03</td>
<td>0.81±0.02</td>
</tr>
<tr>
<td>4</td>
<td>Placebo</td>
<td>4.03±0.01</td>
<td>1.80±0.04</td>
<td>2.23±0.08</td>
<td>0.80±0.02</td>
</tr>
<tr>
<td></td>
<td><em>Origanum extract</em></td>
<td>4.26±0.02</td>
<td>1.90±0.05</td>
<td>2.36±0.04</td>
<td>0.80±0.02</td>
</tr>
<tr>
<td>6</td>
<td>Placebo</td>
<td>3.40±0.02</td>
<td>1.53±0.06</td>
<td>1.87±0.01</td>
<td>0.81±0.01</td>
</tr>
<tr>
<td></td>
<td><em>Origanum extract</em></td>
<td>3.80±0.04</td>
<td>1.73±0.03</td>
<td>2.07±0.06</td>
<td>0.83±0.01</td>
</tr>
<tr>
<td>8</td>
<td>Placebo</td>
<td>3.93±0.01</td>
<td>1.75±0.06</td>
<td>2.18±0.03</td>
<td>0.80±0.01</td>
</tr>
<tr>
<td></td>
<td><em>Origanum extract</em></td>
<td>4.19±0.02</td>
<td>1.89±0.05</td>
<td>2.30±0.05</td>
<td>0.82±0.01</td>
</tr>
</tbody>
</table>

Data are expressed as mean±SE (n=15). *: P<0.05 compared with the placebo at the end of identical every two weeks.

**Immunological analysis**

**Respiratory burst activity**

The respiratory burst activity significantly (p<0.05) enhanced in fish fed with 1% of *Origanum vulgare* extract supplementation feed at the end of the identical every two weeks after feeding in compared to placebo group (Figure 1).

**Phagocytic activity**

Phagocytic activity of blood leucocytes significantly (p<0.05) enhanced in fish treated with 1% of *Origanum vulgare* extract supplementation feed at the end of the identical every two weeks after feeding in compared to placebo group (Figure 2).

**Lysozyme activity**

Lysozyme activity significantly (p<0.05) enhanced in fish fed with 1% of *Origanum vulgare* extract supplementation feed at the end of the identical every two weeks after feeding in compared to placebo group (Figure 3).
Oncorhynchus mykiss extract with optimal dose, and extract [23], and study, that it is in agreement many of studies with dietary phagocytes stimulated by foreign agents [25]. An enhancement in fish, which is a measure of the increase of oxidation level in considered as an important indicator of non-specific defense humoral elements in the serum. Respiratory burst activity is vulgaris immune response of fish. So, this study revealed that are likely to be a result of the enhancement of the non-specific albumin and globulin levels suggest that high concentrations of the non-specific immune system, measurable total protein, coggyria [21], lipopolysaccharide [22], were reported in rainbow trout fed with garlic [20], ginger Hb [19] among the groups.

Discussion

The present study projects the impact of dried Origanum vulgare extract on the hematological and immunological responses in rainbow trout (Oncorhynchus mykiss). The hematological parameters in the present investigation such as RBC, WBC, differential leukocytes counts, hemoglobin, hematocrit, the all of the values of red blood cell indices (MCH, MCHC and MCV) were no significant differences at the end of none of the identical every two weeks after feeding when compared to control group. These observations are in agreement with the obtained results of other researchers, who reported that rainbow trout treated with dietary Aloe vera supplementation were no significant differences in RBC and Hct [18], or RBC and Hb [19] among the groups.

In the present study, the dietary Origanum vulgare extract supplementation enhanced total plasma protein, albumin and globulin values in comparison with control group. Similar results were reported in rainbow trout fed with garlic [20], ginger [21], lipopolysaccharide [22], Laurus nobilis [23], and Coggyria coggyria [24]. Serum proteins are various humoral elements of the non-specific immune system, measurable total protein, albumin and globulin levels suggest that high concentrations are likely to be a result of the enhancement of the non-specific immune response of fish. So, this study revealed that Origanum vulgare extracts incorporated diets helped to increase the humoral elements in the serum. Respiratory burst activity is considered as an important indicator of non-specific defense in fish, which is a measure of the increase of oxidation level in phagocytes stimulated by foreign agents [25]. An enhancement of respiratory burst activity has been identified in the present study, that it is in agreement many of studies with dietary immunostimulants [23,26]. Respiratory burst and phagocytosis response by phagocytes in blood present a major antibacterial defense mechanism in fish [27]. Phagocytosis is one of the most important processes in fish. The main cells involved in phagocytosis in fish are neutrophils and macrophages. These cells remove bacteria mainly by the production of reactive oxygen species (ROS) during a respiratory burst. In addition, neutrophils possess myeloperoxidase in their cytoplasmic granules, which in the presence of halide and hydrogen peroxidase kills bacteria by halogenations of the bacterial cell wall. Moreover, these cells have lysozymes and other hydrolytic enzymes in their lysosomes [28]. Similarly, macrophages can produce nitric oxide in mammals and can be as potent as antibacterial agents, peroxyxynitrites and hydroxyl groups. Phagocytic activity of leucocytes in rainbow trout was enhanced by dietary dose of powdered ginger rhizome [29,30]. Also, in this study an increasing trend in lysozyme activity has been shown which is in agreement with several reports indicating the role of herbal immunostimulants in enhancing lysozyme activity [31-33]. Lysozyme is a humoral component of the non-specific defense mechanism which has the ability to prevent the growth of bacteria by splitting β-1,4 glycosidic bonds in the peptidoglycan of bacterial cell walls, resulting in bacteriolysis. In conclusion, supplementation of OE in aquaculture diets would be use to enhance non-specific immune system in fish. Therefore, further studies are necessary for effective use of Origanum vulgare extract with optimal dose, suitable duration, and method of administration.

Acknowledgement

The authors would like to thank the Iranian Fisheries Research Organization (IFRO) for the financial support.

Funding

Foundation project supported by Iranian Fisheries Science Research Institute (IFRSRI), Tehran-Iran with grant number: 14-12-12-8913-89153.

References


Figure 3: Serum lysozyme activity of different experimental groups observed on different weeks. Data are expressed as mean±SE. Asterisks indicate significantly different from placebo group in the same week. *p<0.05; **p<0.001.
growth of Fusarium and Penicillium species is isolated from food. Hem

7. Karpouthisis I, Pardali E, Feggou E, Kokkini S, Scouras ZG, Mavragani-TP

Effect of dietary oregano essential oil on performance of chickens and
on iron- and induced lipid oxidation of breast, thigh and abdominal fat


in the hematology laboratory: the microhematocrit determination. Am

(5th edn), CBS Publication and distributors, Delhi, India, pp. 479-480.

12. Wintrobe MM (1933) Variation in size and hemoglobin concentration
of erythrocyte in the blood of various vertebrates. Folia Haematol 51:
32-49.


(Carica papaya) extract against the gastrointestinal nematode, Ancylostoma

15. Secombes CJ (1990) Isolation of salmonid macrophages and analysis
of their killing activity. In: Stolen JS, Anderson DP, Roberston BS,
van Muiswinkel WB (Eds.), Techniques in fish immunology. SOS

the rapid spectrophotometric determination of phagocytosis by fish

17. Parry RM, Chandan RC, Shahnaz KM (1965) A rapid and sensitive assay

(2010) Effects of dietary Aloe vera on some specific and nonspecific
immunity in the common carp (Cyprinus carpio). Int J Vet Res 4: 189-
195.

Effect of dietary supplementation of Melissa Officinalis and Aloe Vera
on hematological traits, lipid oxidation of carcass and performance
in rainbow trout (Oncorhynchus mykiss). Online J Animal Feed Res 1: 1-5.

20. Nya EJ (2009) Studies on dietary supplements for the control of
Aeromonas hydrophila infection in rainbow trout (Oncorhynchus
mykiss, Walbaum). Doctor of Philosophy thesis. School of Life Sciences,
Heriot Watt University, Edinburgh, UK.

Aeromonas hydrophila infection in rainbow trout, Oncorhynchus mykiss

22. Nya EJ, Austin B (2009) Use of dietary ginger, Zingiber officinale Roscoe,
as an immunostimulant to control Aeromonas hydrophila infections
in rainbow trout, Oncorhynchus mykiss (Walbaum). J Fish Dis 32(1): 971-
977.

non-specific immune responses of rainbow trout (Oncorhynchus mykiss,

coggaria on rainbow trout (Oncorhynchus mykiss). Fish Shellfish

Humoral and non-specific immune responses in rainbow trout

herbs (Astragalus radix and Gonderaer lucidum) enhance immune
response of carp, Cyprinus carpio and protection against Aeromonas

supplementation diets on hematology and innate immunity in goldfish


(Zingiber officinale) on the haematological and immunological
parameters of rainbow trout Oncorhynchus mykiss. JMPHTR 1: 8-12.

aspera on the immunity and survival of Labeo rohita infected with

mistletoe to enhances cellular non-specific immune responses and
survival of Japanese eel (Anguilla japonica). Fish Shellfish Immunol 24:
67-73.

The effects of different levels of Aloe vera extract on some of the
hematological and non-specific immune parameters in Siberian
sturgeon (Acipenser baerii). Iranian Journal of Fisheries Sciences
16(4): 1234-1247.