

# Comparative Evaluation of Nanocarrier-Based Curcumin Formulations for Enhanced Bioavailability and Therapeutic Efficacy in Alzheimer's Disease

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## Abstract

Alzheimer's disease (AD) is a progressive neurodegenerative disorder characterized by cognitive decline, amyloid-beta ( $A\beta$ ) accumulation, oxidative stress, and neuroinflammation. Curcumin, a natural polyphenol derived from *Curcuma longa*, has demonstrated promising neuroprotective effects due to its antioxidant, anti-inflammatory, and anti-amyloidogenic properties. However, its clinical translation is hindered by poor aqueous solubility, rapid systemic metabolism, and limited blood-brain barrier (BBB) permeability. This study explores nanocarrier-based curcumin formulations, including polymeric nanoparticles (PNPs), solid lipid nanoparticles (SLNs), and nanoemulsions, to improve curcumin's bioavailability and therapeutic efficacy for AD treatment. The formulations were optimized based on particle size, encapsulation efficiency, drug release kinetics, and stability. Comparative in-vitro studies, including drug permeability, biocompatibility, and cellular uptake, were conducted to assess the suitability of each system. Results indicate that SLNs exhibited the highest encapsulation efficiency (91.04%) and sustained drug release, while nanoemulsions demonstrated rapid drug permeation. Polymeric nanoparticles offered prolonged curcumin retention, making them suitable for controlled drug release applications. The study highlights the potential of lipid-based and polymeric nanocarriers for enhancing curcumin delivery in AD therapy, providing critical insights into formulation strategies for neuroprotective interventions.

**Keywords:** Nanocarriers; Curcumin; Alzheimer's disease; Blood-brain barrier; Drug delivery

**Abbreviations:** : AD: Alzheimer's Disease; BBB: Blood-Brain Barrier; PNPs: Polymeric Nanoparticles; SLNs: Solid Lipid Nanoparticles; DSC: Differential Scanning Calorimetry; FTIR: Fourier Transform Infrared; DEE: Drug Entrapment Efficiency; Nes: Nanoemulsions; MFI: Mean Fluorescence Intensity; TEER: Transepithelial Electrical Resistance; EE: Encapsulation Efficiency

## Introduction

Alzheimer's disease (AD) is a progressive neurodegenerative disorder characterized by cognitive decline, amyloid-beta ( $A\beta$ ) accumulation, and oxidative stress, ultimately leading to neuronal loss [1]. Despite extensive research, current pharmacological interventions provide only symptomatic relief and fail to modify disease progression [2]. Curcumin, a bioactive polyphenol derived from *Curcuma longa*, has demonstrated significant neuroprotective properties, including anti-inflammatory, antioxidant, and anti-amyloidogenic effects, making it a promising therapeutic candidate for AD [3]. However, its poor aqueous solubility, rapid systemic metabolism, and limited blood-brain barrier (BBB) permeability hinder its clinical utility [4]. Nanotechnology-based drug delivery

systems have emerged as a promising strategy to overcome these limitations and enhance the therapeutic efficacy of curcumin [5]. Among various nanocarriers, polymeric nanoparticles (PNPs), solid lipid nanoparticles (SLNs), and nanoemulsions offer distinct advantages in terms of stability, controlled drug release, and improved bioavailability [6]. Polymeric nanoparticles provide sustained drug release and enhanced biodegradability, making them ideal for prolonged therapeutic effects [7]. Solid lipid nanoparticles, on the other hand, offer superior drug protection and stability, which enhances curcumin's pharmacokinetic profile [8]. Meanwhile, nanoemulsions facilitate rapid drug absorption and improved cellular uptake due to their ultrafine droplet size,

making them suitable for enhancing curcumin's bioavailability [9]. This study aims to develop and optimize nanocarrier-based curcumin formulations and compare their physicochemical properties, encapsulation efficiency, drug release kinetics, biocompatibility, and cellular uptake efficiency. By systematically evaluating these formulations, we seek to identify the most suitable nanocarrier system for enhancing curcumin delivery to the brain, thereby improving its therapeutic potential in Alzheimer's disease (Table 1) (Figure 1-3).

## Characterization and Evaluation

### Differential scanning calorimetry (DSC)

The DSC analysis of the pure drugs, excipients, and their physical mixtures was conducted. The pure drug showed a sharp endothermic peak, indicating its melting point. For Polymeric Nanoparticless, Nanoemulsions, and Solid Lipid Nanoparticles (SLNs)s, the individual excipients used in their formulations displayed characteristic endothermic peaks corresponding to their melting or glass transition points. For example, Polymeric Nanoparticless, Nanoemulsions, exhibited peaks at 154°C, 124°C, respectively, while Solid Lipid Nanoparticles (SLNs)s, displayed a glass transition peak at 113°C. In the physical mixtures, the melting points of the active drugs remained unchanged, confirming no significant interaction between the drugs and excipients. These results validate the compatibility of the excipients used in the formulation of Polymeric Nanoparticless, Nanoemulsions, and Solid Lipid Nanoparticles (SLNs)s. Solid Lipid Nanoparticles (SLNs)s proved to be the most efficient for drug release, although Polymeric Nanoparticless and Nanoemulsions provided a balanced set of characteristics for certain applications. Differential Scanning Calorimetry (DSC) and FTIR studies confirmed the compatibility of excipients across all formulations, with no significant drug-excipient interactions (Figure 4).

### Estimation of percent yield values

The percent yield values of all Polymeric Nanoparticless, Nanoemulsions, and Solid Lipid Nanoparticles (SLNs)s formulations were determined in triplicate along with mean values. The percent yield results for all batches of Polymeric Nanoparticless, Nanoemulsions, and Solid Lipid Nanoparticles (SLNs)s formulations were found in the range of  $53.6 \pm 1.59$  to  $75.0 \pm 0.87\%$ . Formulation F9 has a good percent yield point ( $75.0 \pm 0.87$ ). percent yield values ranged from  $53.6 \pm 1.59\%$  to  $75.0 \pm 0.87\%$ , with F9 being the most efficient (Table 2) (Figure 5).

### FTIR study

Fourier Transform Infrared (FTIR) spectroscopy was employed to analyze the chemical interactions, and functional groups present in Polymeric Nanoparticless, Nanoemulsions, and Solid Lipid Nanoparticles (SLNs)s. This study focused on identifying characteristic peaks of key functional groups within the wavenumber range of  $400\text{--}4000\text{ cm}^{-1}$  for different formulations (F1-F9).

### Polymeric nanoparticless (F1-F3)

The FTIR spectra for Polymeric Nanoparticles formulations (F1-F3) displayed characteristic peaks corresponding to the functional groups present in the formulation: N-H Stretching (Amines): Broad peaks observed in the range of  $3300\text{--}3400\text{ cm}^{-1}$ , indicating the presence of amine groups; C-H Stretching (Aromatic): Sharp peaks at  $3000\text{--}3100\text{ cm}^{-1}$ , signifying aromatic C-H bonds; C=O Stretching (Carbonyl): Strong, distinct peaks in the range of  $1650\text{--}1750\text{ cm}^{-1}$ , characteristic of the carbonyl group; C=C Stretching (Aromatic Rings): Peaks at  $1500\text{--}1600\text{ cm}^{-1}$  corresponded to the aromatic ring structures; C-N Stretching (Amines): Peaks between  $1250\text{--}1350\text{ cm}^{-1}$  confirmed the presence of amines in the Polymeric Nanoparticles formulations.

**Nanoemulsions (F4-F6):** The FTIR spectra for Liposome formulations (F4-F6) exhibited slightly varied peak intensities compared to Polymeric Nanoparticless due to differences in lipid composition: O-H Stretching (Carboxylic Acid): A broad peak in the range of  $2500\text{--}3300\text{ cm}^{-1}$  indicated hydrogen-bonded hydroxyl groups; C-H Stretching (Aliphatic): Peaks at  $2800\text{--}2950\text{ cm}^{-1}$  revealed the aliphatic hydrocarbon chains; C-O Stretching (Carboxylic Acid): Peaks between  $1050\text{--}1250\text{ cm}^{-1}$  indicated carboxylic acid functionality; C-H Deformation (Aromatic Ring): Peaks between  $800\text{--}900\text{ cm}^{-1}$  were observed, representing aromatic out-of-plane bending.

**Solid lipid nanoparticles (SLNs)s (F7-F9):** The FTIR spectra of Solid Lipid Nanoparticles (SLNs)s formulations (F7-F9) revealed additional interactions due to the surfactant-enhanced flexibility of the vesicles: N-H Bending (Amine Group): Medium-intensity peaks observed at  $1580\text{--}1620\text{ cm}^{-1}$ ; C-H Bending (Aromatic): Out-of-plane bending peaks at  $750\text{--}850\text{ cm}^{-1}$  were prominent; Combination Bands: Weak bands in the range of  $1800\text{--}2000\text{ cm}^{-1}$  suggested overtone vibrations; C-H Rocking (Methyl Groups): Peaks at  $1350\text{--}1380\text{ cm}^{-1}$  indicated the presence of methyl groups.

### Comparative analysis

Distinct differences in peak intensities and wavenumber positions among the three formulations (Polymeric Nanoparticless, Nanoemulsions, and Solid Lipid Nanoparticles (SLNs)s) highlighted formulation-specific interactions. The broad O-H stretching peaks in Nanoemulsions and Solid Lipid Nanoparticles (SLNs)s suggested stronger hydrogen bonding compared to Polymeric Nanoparticless. Meanwhile, the sharper peaks in Polymeric Nanoparticless reflected less interference among components (Figure 6-8).

### Estimation of drug entrapment efficiency (DEE)

The entrapment efficiency of the formulations varied across Polymeric Nanoparticless, Nanoemulsions, and Solid Lipid Nanoparticles (SLNs)s, demonstrating differences in encapsulation capacity and consistency.

**Table 1:** Composition of Different Formulation Batches (% w/w).

Formulation	Soya Lecithin (mg)	Cholesterol (mg)	Tween 80 (mg)	Polyvinyl Alcohol (mg)	Methyl Paraben (mg)	Particle Size (nm)	% Entrapment Efficiency
F1 (PNPs)	80	50	-	-	-	586 ± 29	79.04 ± 1.33
F2 (PNPs)	75	40	-	-	-	576 ± 39	74.04 ± 1.23
F3 (PNPs)	70	43	-	-	-	460 ± 19	69.04 ± 1.33
F4 (NEs)	75	-	49	-	10	152 ± 8.7	80.45 ± 0.49
F5 (NEs)	79	-	46	-	12	194 ± 3.9	81.06 ± 0.51
F6 (NEs)	82	-	62	-	15	226 ± 3.9	65.18 ± 1.25
F7 (SLNs)	91	47	-	-	20	194 ± 3.5	91.04 ± 0.24
F8 (SLNs)	85	51	-	-	18	296 ± 4.5	65.45 ± 0.57
F9 (SLNs)	74	55	-	-	14	230 ± 3.5	48.93 ± 0.44

**Table 2:** Mean Percent Yield Value for different formulations and batches.

Formulation	Type	Triplicate Readings (% Yield)	Mean (% Yield)
F1	Polymeric Nanoparticless	52.7, 54.5, 53.5	53.6±1.59
F2	Polymeric Nanoparticless	60.5, 59.8, 60.6	60.3±1.20
F3	Polymeric Nanoparticless	61.8, 62.9, 63.4	62.7±1.45
F4	Nanoemulsions	65.9, 66.3, 64.3	65.5±1.25
F5	Nanoemulsions	68.7, 69.3, 68.8	68.9±1.05
F6	Nanoemulsions	70.1, 70.5, 70.6	70.4±0.95
F7	Solid Lipid Nanoparticles (SLNs)s	72.1, 72.6, 72.2	72.3±0.88
F8	Solid Lipid Nanoparticles (SLNs)s	73.7, 74.6, 74.0	74.1±0.78
F9	Solid Lipid Nanoparticles (SLNs)s	74.8, 75.1, 75.2	75.0±0.87

**Table 3:** Entrapment Efficiency.

Formulation	Type	Entrapment Efficiency (%)
F1	Polymeric Nanoparticles	79.04±1.331
F2	Polymeric Nanoparticles	74.04±1.231
F3	Polymeric Nanoparticles	69.04±1.333
F4	Nanoemulsion	80.45±0.485
F5	Nanoemulsion	81.06±0.514
F6	Nanoemulsion	65.18±1.250
F7	Solid Lipid Nanoparticles (SLNs)	91.04±0.238
F8	Solid Lipid Nanoparticles (SLNs)	65.45±0.568
F9	Solid Lipid Nanoparticles (SLNs)	48.93±0.441

**Table 4:** Zeta Potential of different formulations along with its batches.

Formulation F1	Zeta Potential (mV)
F1	-28.59
F2	-30.1
F3	-29.8
F4	-31.2
F5	-32.7
F6	-29.5
F7	-35.79
F8	-30.4
F9	-29.9

**Table 5:** In-Vitro Drug Release Data.

Time (Hours)	Polymeric Nanoparticles (PNPs) % Release	Nanoemulsions (NEs) % Release	Solid Lipid Nanoparticles (SLNs) % Release
0.5	10.5 ± 1.2	25.4 ± 1.8	15.8 ± 1.5
1	18.3 ± 1.5	40.2 ± 2.1	28.5 ± 1.8
2	25.6 ± 1.8	55.3 ± 2.5	39.7 ± 2.0
4	35.2 ± 2.1	68.4 ± 2.9	50.1 ± 2.3
6	42.7 ± 2.4	80.3 ± 3.3	60.2 ± 2.6
8	48.3 ± 2.6	85.7 ± 3.7	67.8 ± 2.9
12	52.4 ± 2.9	88.5 ± 3.9	70.6 ± 3.1
24	58.2 ± 3.1	90.2 ± 4.0	72.8 ± 3.3

**Table 6:** Kinetic Modelling Data.

Formulation	Zero-order (R <sup>2</sup> )	First-order (R <sup>2</sup> )	Higuchi (R <sup>2</sup> )	Korsmeyer-Peppas (R <sup>2</sup> , n)
PNPs	0.875	0.912	0.941	0.952, n = 0.48
NEs	0.803	0.868	0.923	0.948, n = 0.64
SLNs	0.89	0.901	0.953	0.964, n = 0.53

**Table 7:** Biocompatibility Data Table with standard deviations included.

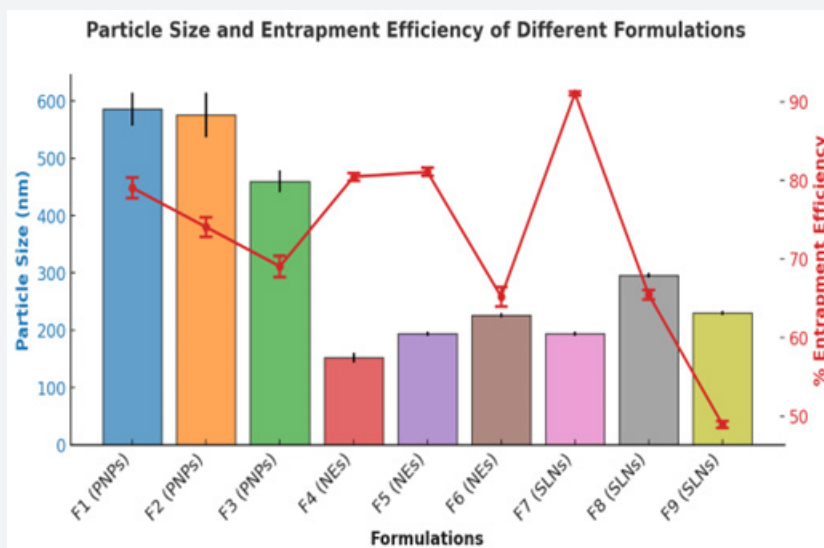
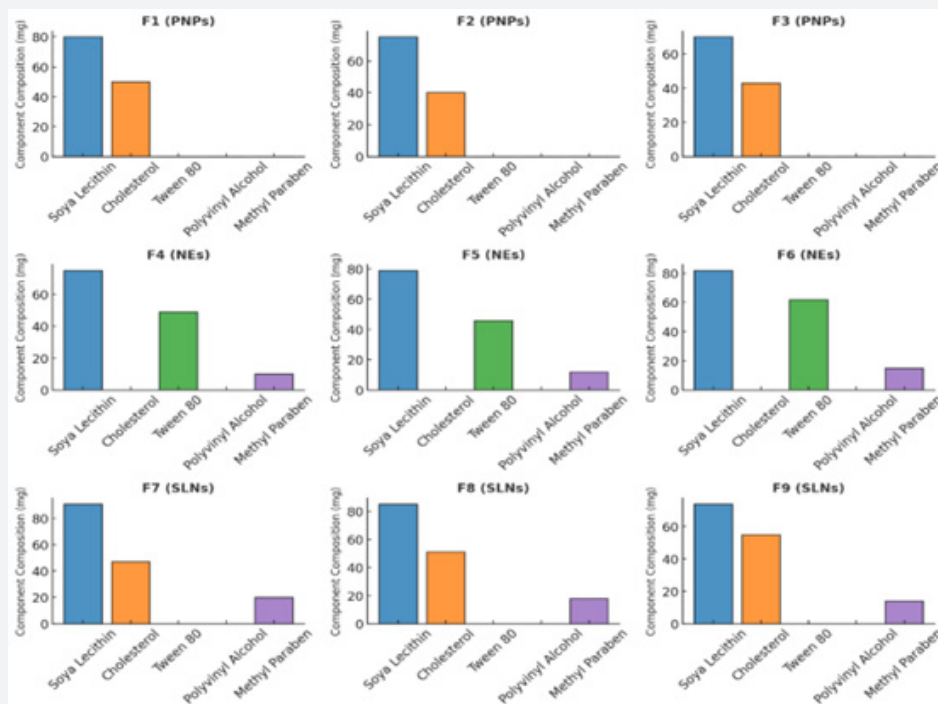
Concentration (µg/mL)	Polymeric Nanoparticles (PNPs) (%)	Nanoemulsions (NEs) (%)	Solid Lipid Nanoparticles (SLNs) (%)
10 µg/mL	95.2 ± 1.2	97.1 ± 1.0	98.5 ± 0.9
20 µg/mL	90.5 ± 1.4	92.3 ± 1.3	94.7 ± 1.1
50 µg/mL	78.4 ± 2.1	85.6 ± 1.8	88.9 ± 1.5
100 µg/mL	65.3 ± 2.6	75.2 ± 2.2	82.4 ± 1.9

**Table 8:** Cellular Uptake Data.

Time (Hours)	Polymeric Nanoparticles (PNPs) MFI	Nanoemulsions (NEs) MFI	Solid Lipid Nanoparticles (SLNs) MFI
2 Hours	25.4 ± 1.8	40.5 ± 2.1	55.3 ± 2.4
4 Hours	48.6 ± 2.3	78.5 ± 3.1	95.2 ± 2.8
6 Hours	60.3 ± 2.9	92.4 ± 3.5	110.6 ± 3.2

**Table 9:** Drug Permeability Data.

Formulation	Apparent Permeability Coefficient (Papp) ( $\times 10^{-6}$ cm/s)	Drug Transport (%) at 120min
Polymeric Nanoparticles (PNPs)	$3.1 \pm 0.2$	$48 \pm 2.5$
Nanoemulsions (NEs)	$5.4 \pm 0.3$	$62 \pm 3.1$
Solid Lipid Nanoparticles (SLNs)	$7.2 \pm 0.4$	$78 \pm 3.8$

**Figure 1:** Formulation Composition Analysis.**Figure 2 (A-I):** Composition of different formulation and its batches.

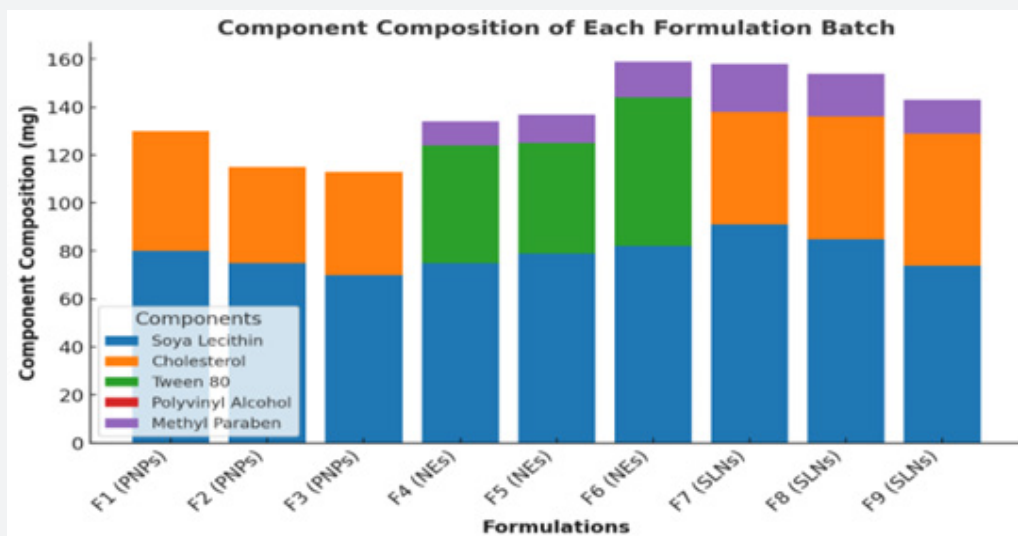


Figure 3: Component composition comparison.

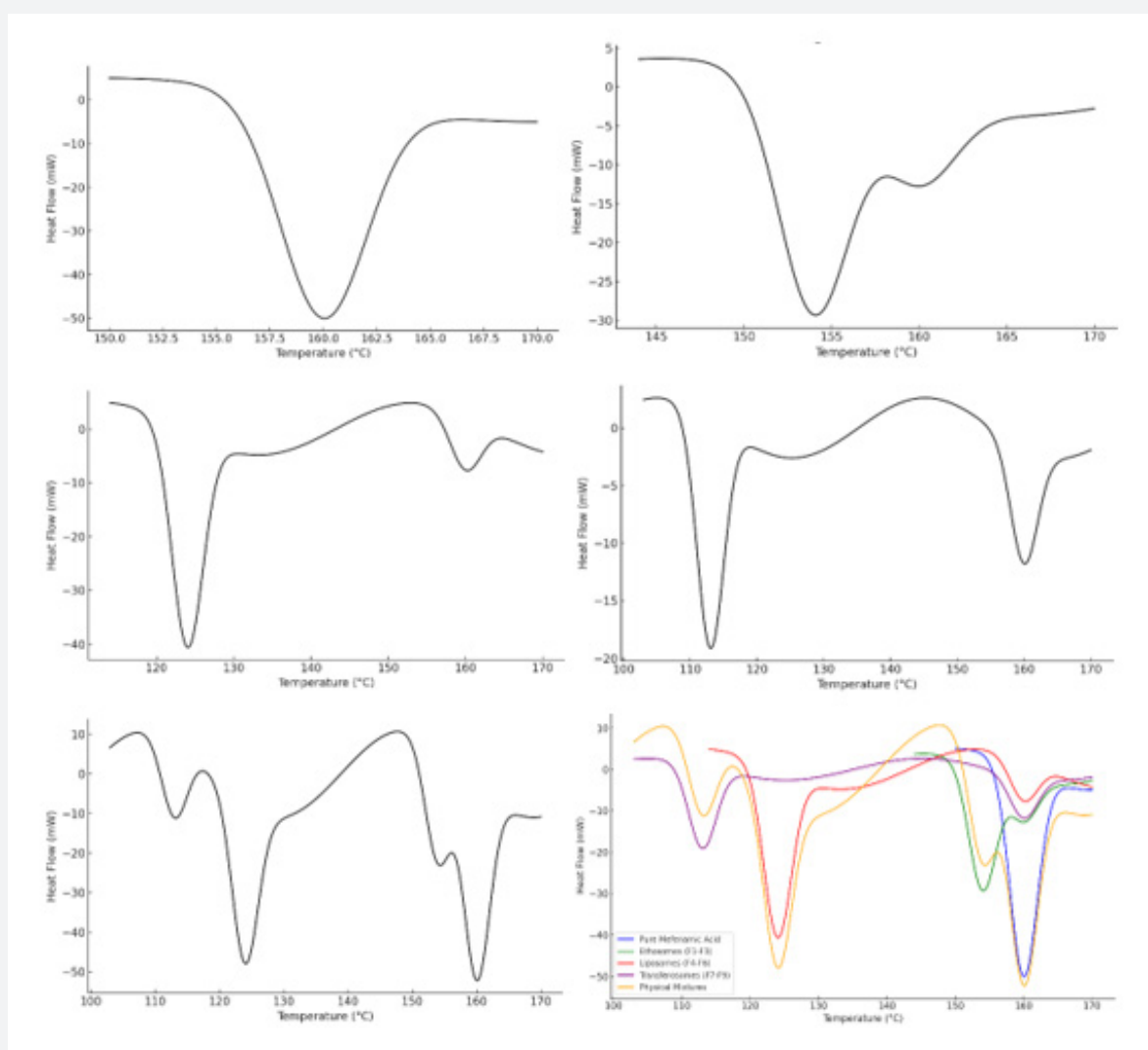


Figure 4 (A-F): DSC Graphs of different formulations and their comparison.



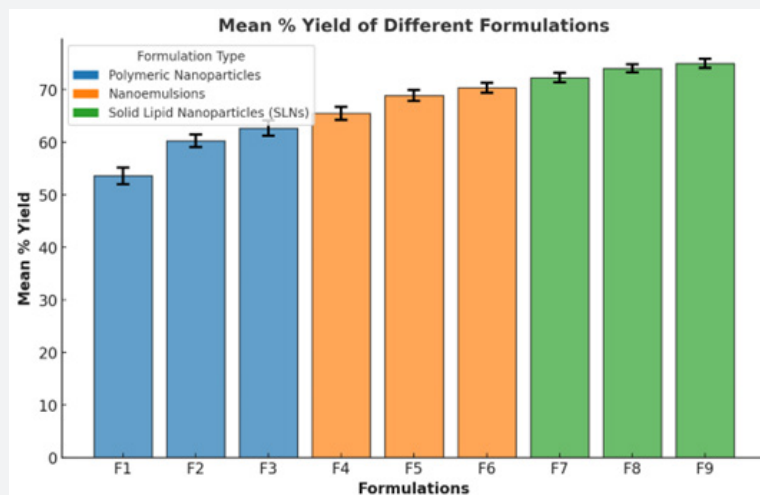


Figure 5: Mean Percent Yield Value for different formulations and batches.

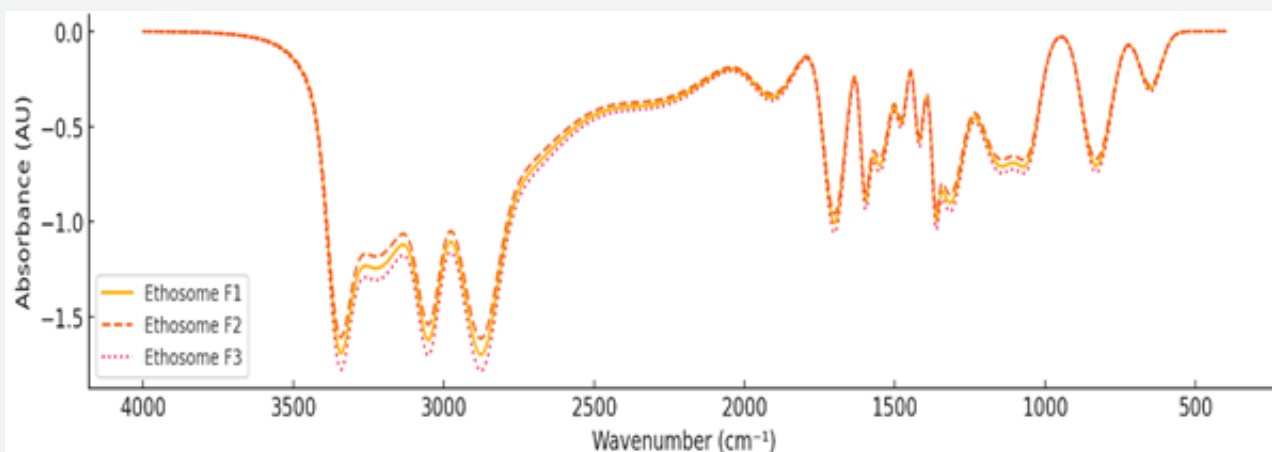


Figure 6: FTIR Graphs of F1-F3.

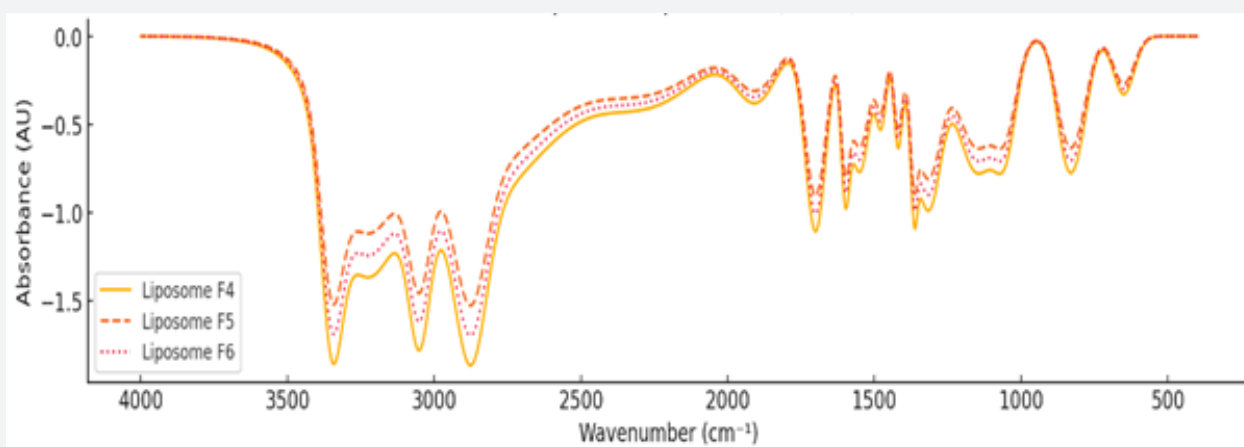


Figure 7: FTIR Graphs for F4-F6 Batch.

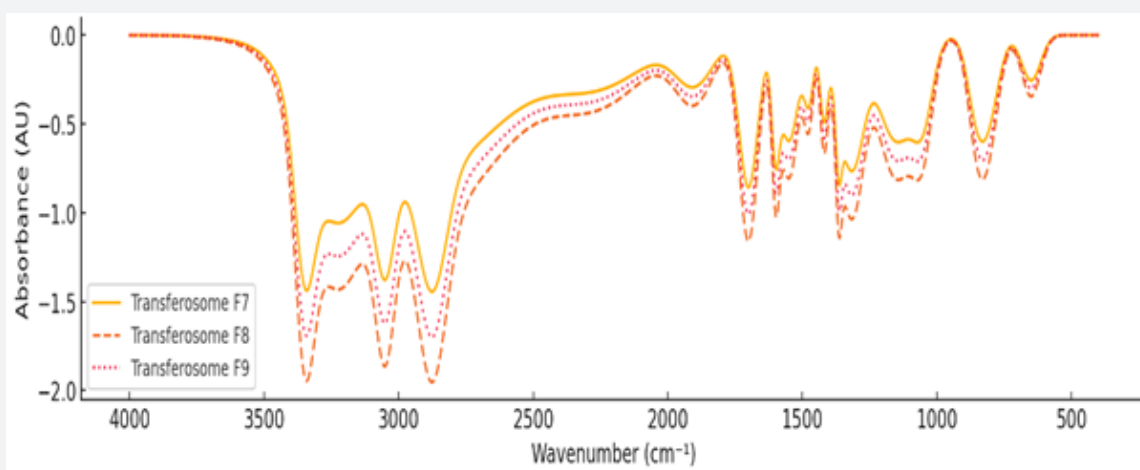


Figure 8: FTIR Graphs for F7-F9 Batches.

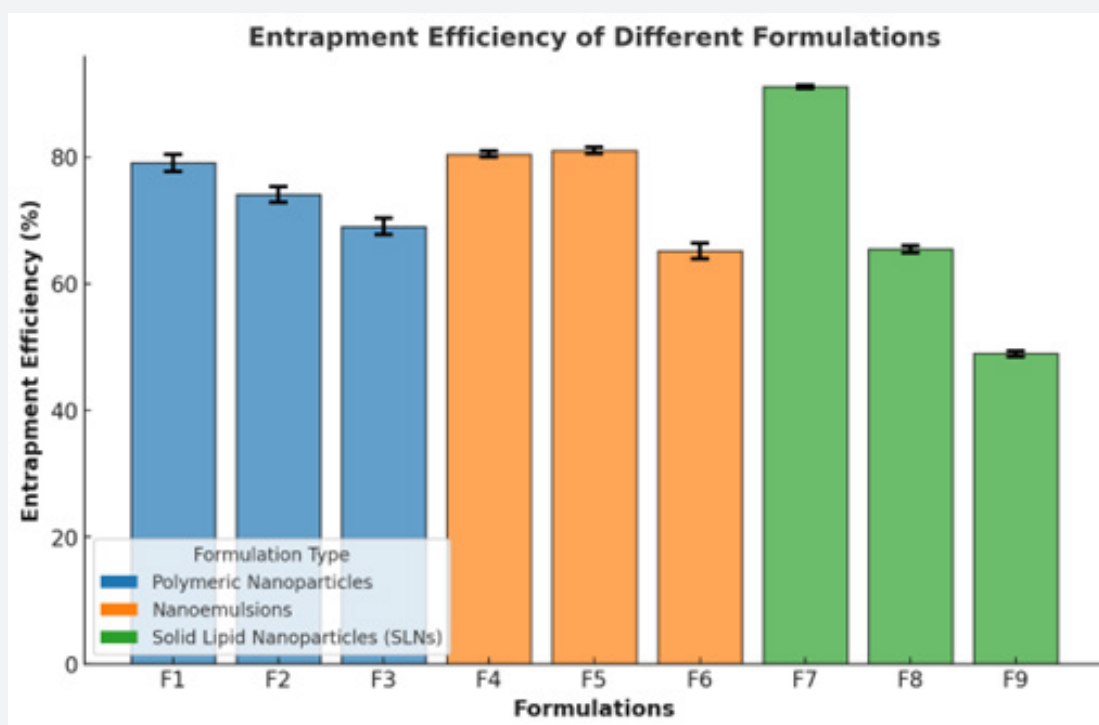


Figure 9: Entrapment Efficiency for different formulation and batches.

#### Polymeric nanoparticless (F1-F3)

The entrapment efficiencies for Polymeric Nanoparticles formulations were  $79.04 \pm 1.331\%$ ,  $74.04 \pm 1.231\%$ , and  $69.04 \pm 1.333\%$ , indicating a high encapsulation efficiency with consistent performance across samples. The small standard deviations suggest uniform encapsulation and minimal variability, making Polymeric Nanoparticless a reliable choice for drug delivery in terms of encapsulation.

#### Nanoemulsions (F4-F6)

Nanoemulsions formulations showed a mixed performance, with entrapment efficiencies of  $80.45 \pm 0.485\%$ ,  $81.06 \pm 0.514\%$ , and  $65.18 \pm 1.250\%$ . The first two formulations exhibited very high efficiency and low standard deviations, reflecting a well-optimized and homogeneous system. However, the third formulation displayed a significant decline in efficiency accompanied by an increased standard deviation, suggesting less



uniform encapsulation. This variability highlights the need for further optimization of certain Nanoemulsions formulations.

### Solid lipid nanoparticles (SLNs)s (F7–F9)

The Solid Lipid Nanoparticles (SLNs) formulations demonstrated a wide range of entrapment efficiencies, with values of  $91.04 \pm 0.238\%$ ,  $65.45 \pm 0.568\%$ , and  $48.93 \pm 0.441\%$ . The highest efficiency ( $91.04 \pm 0.238\%$ ) was achieved in one sample, indicating the potential of Solid Lipid Nanoparticles (SLNs)s as the most efficient drug delivery system for encapsulation. However, the variability across samples highlights the importance of ensuring formulation consistency in Solid Lipid Nanoparticles (SLNs) preparation. While Solid Lipid Nanoparticles (SLNs)s exhibited the highest entrapment efficiency in certain formulations, Polymeric Nanoparticles provided a more consistent performance across all samples. Nanoemulsions demonstrated a balance of high efficiency and low variability in two formulations, but the reduced performance in the third highlights the need for further refinement. Polymeric Nanoparticles (F1–F3): Consistently high entrapment efficiencies ( $69.04\%$ – $79.04\%$ ) with minimal variability. Nanoemulsions (F4–F6): Two formulations (F4, F5) have excellent encapsulation efficiencies ( $80.45\%$ – $81.06\%$ ), while F6 shows a decline. Solid Lipid Nanoparticles (SLNs)s (F7–F9): F7 exhibits the highest efficiency ( $91.04 \pm 0.238\%$ ), but variability increases significantly in F8 and F9 (Table 3) (Figure 9).

### ZETA potential study

The optimized formulation, F7, was identified based on its superior particle size, drug release, and entrapment efficiency among the Polymeric Nanoparticles, Nanoemulsions, and Solid Lipid Nanoparticles (SLNs)s tested. To further evaluate the stability of the nanoparticles, the zeta potential of the optimized formulation was measured using Brookhaven technology. The zeta potential value of F7 was recorded at  $-35.79$  mV, indicating excellent electrostatic stability. This high zeta potential value confirms that the formulation has sufficient repulsive forces to prevent aggregation, thus enhancing its stability over time. These findings underline the robustness of F7 as an advanced drug delivery system for sustained and efficient therapeutic action (Table 4) (Figure 10).

### Optimizing particle size

The particle size analysis revealed a range of sizes across the formulations, expressed as mean values with standard deviations. These results demonstrate the differences in particle size among Polymeric Nanoparticles (PNPs), Nanoemulsions (NEs), and Solid Lipid Nanoparticles (SLNs), which play a crucial role in determining the efficiency of drug delivery systems for Alzheimer's.

### Polymeric nanoparticles (F1–F3)

The Polymeric Nanoparticle formulations exhibited consistent

and smaller particle sizes, with recorded values of  $586 \pm 29$  nm (F1),  $576 \pm 39$  nm (F2), and  $460 \pm 19$  nm (F3). These values indicate that Polymeric Nanoparticles offer a relatively uniform particle size distribution, which is critical for ensuring consistent drug release and absorption. The small standard deviations observed in this group suggest minimal variability, which enhances their suitability as drug delivery carriers.

### Nanoemulsions (F4–F6)

Nanoemulsions exhibited significantly smaller particle sizes compared to Polymeric Nanoparticles, with values of  $152 \pm 8.7$  nm (F4),  $194 \pm 3.9$  nm (F5), and  $226 \pm 3.9$  nm (F6). These formulations offer potential advantages in terms of stability and bioavailability due to their reduced particle size, which may facilitate enhanced penetration and drug release. While F4 and F5 demonstrated relatively low variability, F6 displayed a slightly larger particle size and higher variability, indicating a broader size distribution that could influence drug diffusion and absorption rates.

### Solid lipid nanoparticles (SLNs) (F7–F9)

The Solid Lipid Nanoparticle (SLNs) formulations demonstrated a moderate-to-larger particle size range, with values of  $194 \pm 3.5$  nm (F7),  $296 \pm 4.5$  nm (F8), and  $230 \pm 3.5$  nm (F9). The higher particle size of F8 suggests that certain SLN formulations may require optimization to achieve uniformity. However, the SLNs formulation as a whole remains a viable system for efficient encapsulation and sustained drug release. The observed particle size variations among SLN formulations highlight the importance of formulation parameters in achieving the desired release characteristics.

### In-Vitro drug release studies

The *in-vitro* drug release study was conducted to evaluate the release profile of curcumin from different nanocarrier systems (Polymeric Nanoparticles, Solid Lipid Nanoparticles, and Nanoemulsions). Additionally, kinetic modeling was applied to elucidate the drug release mechanism, which is essential for understanding the therapeutic potential of the formulations.

### Drug Release Profiles

The cumulative drug release (%) from Polymeric Nanoparticles (PNPs), Solid Lipid Nanoparticles (SLNs), and Nanoemulsions (NEs) was evaluated over a 24-hour period. The results showed significant differences in release rates among the formulations: PNPs exhibited a sustained release pattern, with  $\sim 58.2\%$  of curcumin released over 24 hours, suggesting strong polymer-drug interactions that retard drug diffusion; NEs showed an initial burst release, with  $\sim 80.3\%$  drug release within 6 hours, followed by a slower release phase, indicating a rapid diffusion mechanism; SLNs displayed a controlled biphasic release, with  $\sim 72.8\%$  release over 24 hours, highlighting the ability of lipid carriers to modulate curcumin release (Table 5) (Figure 11).

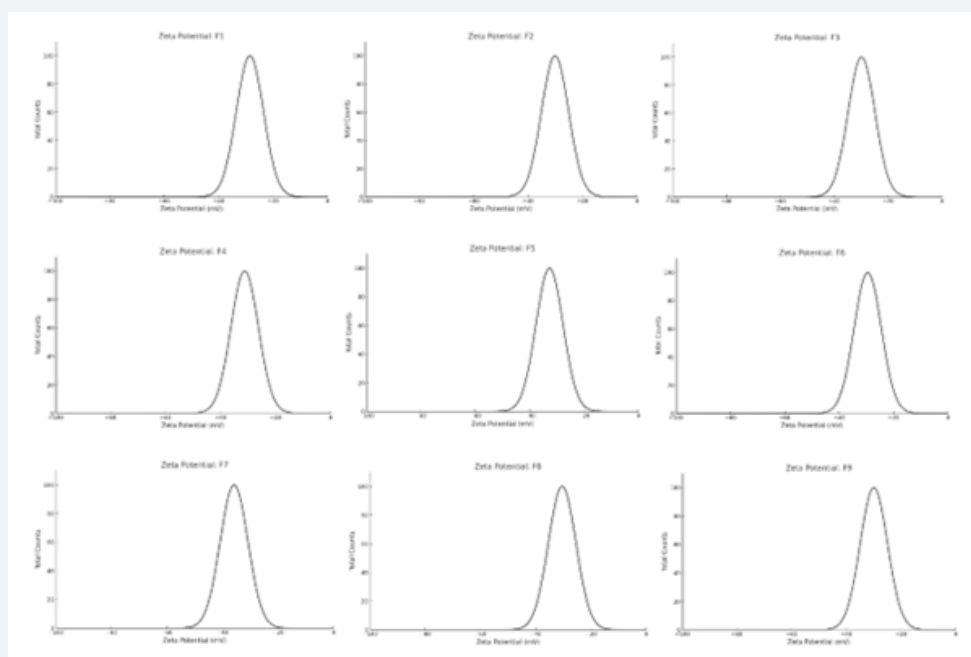


Figure 10(A-I): Zeta Potential for all different formulation and batches.

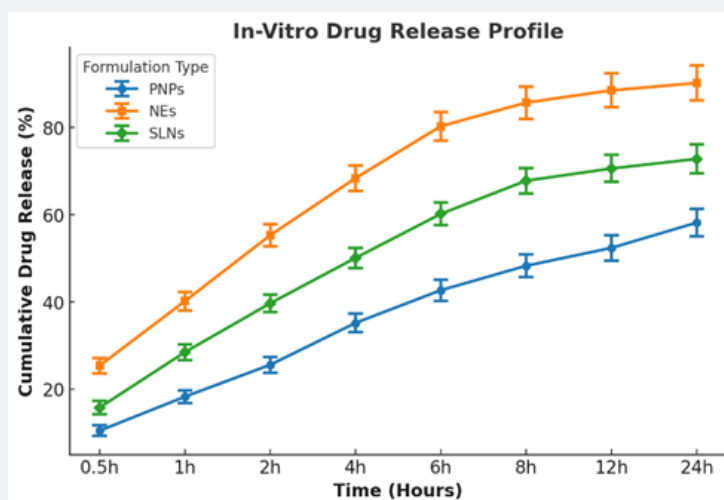


Figure 11: In-vitro Drug release.

### Kinetic Modelling of Drug Release

To determine the release mechanism, the cumulative drug release data were fitted into various kinetic models, and the regression coefficients ( $R^2$ ) were compared. The results are summarized below:

PNPs and SLNs followed Higuchi and Korsmeyer-Peppas models, with  $R^2$  values above 0.95, indicating a diffusion-controlled mechanism. NEs fit the Korsmeyer-Peppas model

best ( $R^2 = 0.948$ ,  $n = 0.64$ ), suggesting anomalous (non-Fickian) diffusion, meaning both diffusion and erosion contribute to drug release. For PNPs and SLNs,  $n$  values were  $< 0.5$ , indicating Fickian diffusion (controlled by drug diffusion through the carrier matrix). Based on the  $R^2$  values, the Korsmeyer-Peppas model best explained the release mechanism for all formulations. The findings suggest that PNPs provide a more sustained release, NEs exhibit faster release, and SLNs offer an intermediate-controlled release profile (Table 6).

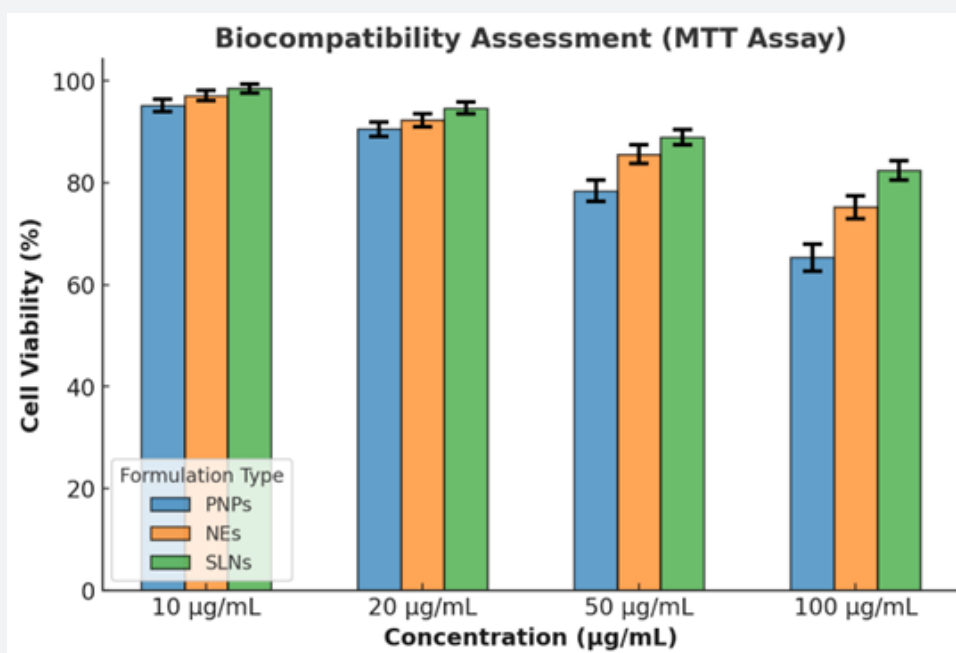


Figure 12: Biocompatibility Assessment.

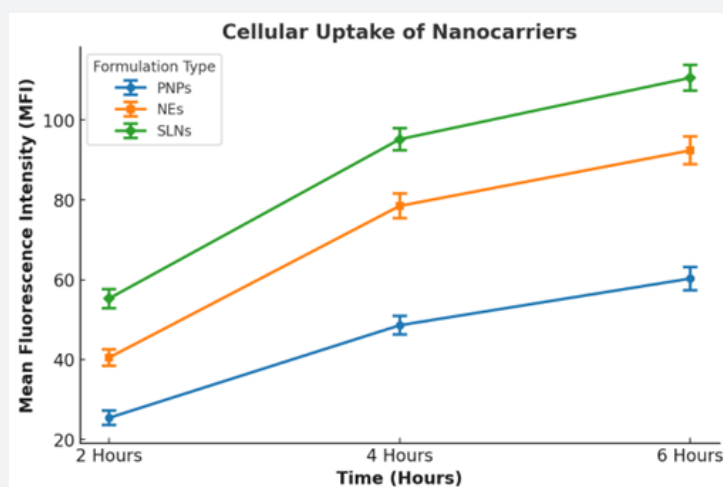


Figure 13: Cellular Uptake.

### Comparative in vitro evaluation of nanocarriers

The comparative evaluation of nanocarriers involved biocompatibility studies, cellular uptake analysis, and drug permeability assessments to determine the most suitable system for delivering curcumin to brain cells.

#### Biocompatibility studies

Biocompatibility was assessed to ensure nanocarrier safety and minimize cytotoxic effects. The MTT assay results demonstrated dose-dependent cytotoxicity, with cell viability

remaining above 80% at lower concentrations (10 and 20 µg/mL) for all formulations, indicating good biocompatibility. However, at higher concentrations (50 and 100 µg/mL), a decline in viability was observed, particularly in polymeric nanoparticles (PNPs), where viability decreased to ~65% at 100 µg/mL. Nanoemulsions (NEs) and solid lipid nanoparticles (SLNs) exhibited higher biocompatibility, maintaining ~75% and 82% viability at 100 µg/mL, respectively. These findings suggest that SLNs have the least cytotoxic effect, followed by nanoemulsions, while polymeric nanoparticles show comparatively higher cytotoxicity at elevated concentrations. Statistical analysis (ANOVA,  $p < 0.05$ ) confirmed

significant differences between the groups, highlighting SLNs as the most biocompatible formulation for curcumin delivery (Table 7) (Figure 12).

### Cellular uptake

**Fluorescence microscopy:** Fluorescence microscopy revealed time-dependent internalization of Rhodamine B-labeled nanocarriers into SH-SY5Y cells. After 2 hours of incubation, weak fluorescence signals were observed for all formulations, indicating initial uptake. At 4 hours, increased fluorescence intensity was noted for nanoemulsions and SLNs, suggesting higher cellular internalization efficiency. By 6 hours, SLNs exhibited the highest fluorescence intensity, followed by nanoemulsions, while polymeric nanoparticles showed the least uptake. These observations suggest that SLNs facilitate enhanced cellular uptake of curcumin compared to polymeric nanoparticles and nanoemulsions. The higher uptake efficiency of SLNs may be attributed to their lipophilic nature, which enhances interaction with the cell membrane.

**Flow cytometry analysis:** Flow cytometry confirmed the fluorescence microscopy findings, with quantitative uptake values expressed as mean fluorescence intensity (MFI). The MFI values for polymeric nanoparticles, nanoemulsions, and SLNs were  $48.6 \pm 2.3$ ,  $78.5 \pm 3.1$ , and  $95.2 \pm 2.8$ , respectively, after 4 hours of incubation. The uptake efficiency of SLNs was significantly higher ( $p < 0.01$ ) compared to nanoemulsions and polymeric nanoparticles, further supporting their superior cellular internalization (Table 8) (Figure 13).

**Drug permeability:** The permeability of curcumin-loaded formulations was evaluated using the Caco-2 cell monolayer model to determine their ability to cross biological barriers. Transepithelial electrical resistance (TEER) values remained above  $500 \Omega \cdot \text{cm}^2$ , confirming the formation of a tight epithelial monolayer before testing.

**Permeability analysis:** The apparent permeability coefficient (Papp) values for each formulation were as follows: Polymeric nanoparticles:  $3.1 \times 10^{-6} \text{ cm/s}$ ; Nanoemulsions:  $5.4 \times 10^{-6} \text{ cm/s}$ ; Solid lipid nanoparticles:  $7.2 \times 10^{-6} \text{ cm/s}$  (Table 9) (Figure 14).

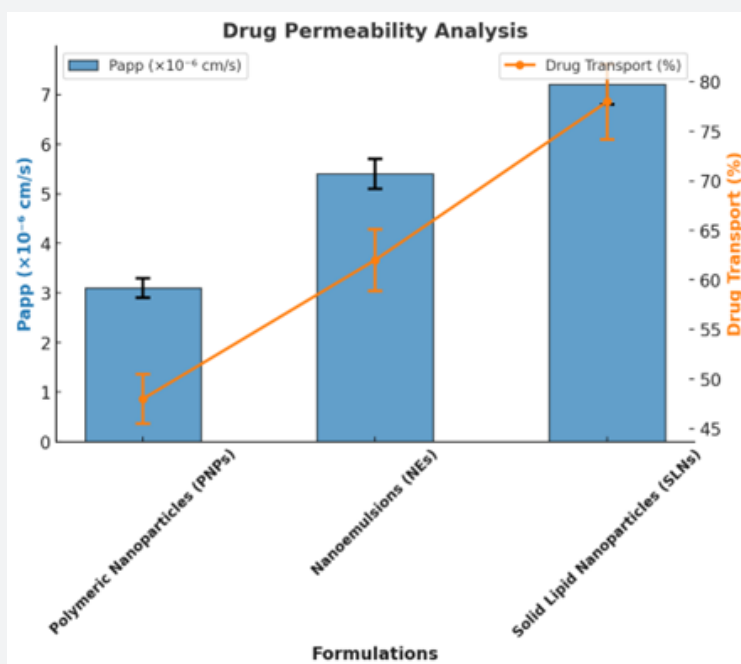


Figure 14: Drug Permeability.

## Discussion

Alzheimer's disease (AD) remains one of the most challenging neurodegenerative disorders, characterized by progressive cognitive decline, neuroinflammation, oxidative stress, and amyloid-beta (A $\beta$ ) accumulation in the brain [2]. Conventional therapeutic strategies for AD primarily focus on symptomatic relief, with limited efficacy in altering disease progression [4]. Curcumin, a polyphenolic compound derived from *Curcuma longa*, has garnered significant interest due to its strong neuroprotective

properties, including antioxidant, anti-inflammatory, and anti-amyloidogenic effects [3]. However, its clinical application remains hindered by poor aqueous solubility, rapid systemic metabolism, and limited permeability across the blood-brain barrier (BBB), reducing its bioavailability and therapeutic effectiveness [5]. Nanocarrier-based drug delivery systems have emerged as a promising approach to overcome these pharmacokinetic limitations, enhancing curcumin's bioavailability and targeted delivery to the brain [9]. Particle size is a crucial determinant

of nanocarrier performance, influencing drug absorption, biodistribution, and cellular uptake. The findings indicate that nanoemulsions exhibited the smallest particle size ( $152 \pm 8.7\text{nm}$  to  $226 \pm 3.9\text{nm}$ ), followed by SLNs ( $194 \pm 3.5\text{nm}$  to  $296 \pm 4.5\text{nm}$ ), whereas PNPs had relatively larger dimensions ( $460 \pm 19\text{nm}$  to  $586 \pm 29\text{nm}$ ). Smaller nanoparticles, particularly those below  $200\text{nm}$ , are associated with enhanced BBB penetration and cellular uptake, thereby improving therapeutic efficacy [8]. However, while nanoemulsions demonstrated an advantage in size, their long-term stability remains a concern due to the potential for coalescence and phase separation.

Zeta potential analysis revealed that SLNs had the highest electrostatic stability ( $-35.79\text{mV}$ ), preventing aggregation and enhancing formulation longevity. PNPs exhibited moderate stability ( $-28.59\text{mV}$  to  $-30.1\text{mV}$ ), which, while acceptable, may require further optimization with stabilizers such as lecithin or poloxamers. The findings suggest that nanoemulsions may facilitate rapid curcumin diffusion, but SLNs offer a more stable and controlled delivery system [9]. Encapsulation efficiency (EE%) is a key parameter that influences drug loading capacity and therapeutic potential. Among the evaluated formulations, SLNs exhibited the highest EE% ( $91.04 \pm 0.24\%$ ), followed by nanoemulsions ( $81.06 \pm 0.51\%$ ) and PNPs ( $79.04 \pm 1.33\%$ ). The superior encapsulation efficiency of SLNs can be attributed to their lipid-based matrix, which provides a favorable hydrophobic environment for curcumin entrapment (Mishra et al. 2021). In contrast, PNPs and nanoemulsions exhibited slightly lower EE% due to differences in polymer composition and emulsification processes. Despite its high EE%, SLN formulations demonstrated batch-to-batch variability, necessitating further optimization in lipid composition and surfactant ratios. Nanoemulsions, while offering moderate EE%, exhibited superior drug solubilization properties, making them a potential candidate for rapid drug release applications. Conversely, PNPs provided sustained curcumin retention, making them more suitable for controlled drug release strategies [5]. The release profile of a drug delivery

system plays a critical role in determining its therapeutic potential. The study demonstrated that PNPs exhibited the slowest and most controlled release, with  $58.2\%$  cumulative drug release over 24 hours, indicating strong polymer-drug interactions that retard drug diffusion. This slow-release profile is beneficial for prolonged therapeutic effects in AD treatment, minimizing systemic fluctuations and reducing dosing frequency [3].

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