

Recurrence of Focal Segmental Glomerulosclerosis (FSGS) in the Renal Allograft; an Update



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Abstract

Patients with primary focal segmental glomerulosclerosis (FSGS) have a high risk of disease recurrence post renal transplantation; this is usually characterised by significant proteinuria and associated with poor graft survival. FSGS recurrence is a challenging medical condition, not only because there is no recognised predictor of FSGS recurrence in the allograft, but also because there are no available international guidelines for treatment of this condition which add more mystery to the dilemma created by this vague syndrome.

Our work aimed to draw the attention to the latest available evidence addressing recurrence of FSGS in the renal allograft, in an attempt to improve our understanding of this condition. This also helps to guide the future studies towards the promising approaches in diagnosis and management of this condition. Ultimately this will result in better graft survival and lower the demands for another renal transplant giving the universal shortage of transplantable organs.

Keywords: Transplantation; FSGS; Recurrence; Columbia classification

Abbreviations: FSGS: Focal Segmental Glomerulosclerosis; NSAIDs: Non-Steroidal Anti-Inflammatory Drugs; ACEI: Angiotensin-Converting Enzyme Inhibitor; ARB: Angiotensin II Receptor Blocker; SuPAR: Soluble Urokinase Receptor

Discussion

FSGS is a histological description of the glomerular pathological findings; Clinically it is characterised by proteinuria, usually of nephrotic range, hypoalbuminaemia and variable degrees of renal impairment [1]. Severe nephrotic syndrome may increase the risk of thromboembolism [2]. FSGS is classified according to the aetiology into primary (idiopathic) and secondary. Secondary FSGS may be due to infection (like HIV and parvovirus B19), drugs and toxins (pamidronate toxicity and heroin), obesity, renal atheroembolic disease and reduced functioning renal mass with hyper filtration of the remaining nephrons [1].

Another important point to be considered is the possibility of podocin gene mutation. The mutation of NPHS2 (encoding for podocin) is one of the commonest causes of the familial nephrotic syndrome. Moreover, there is convincing evidence that it occurs in some patients with sporadic nephrotic syndrome [3].

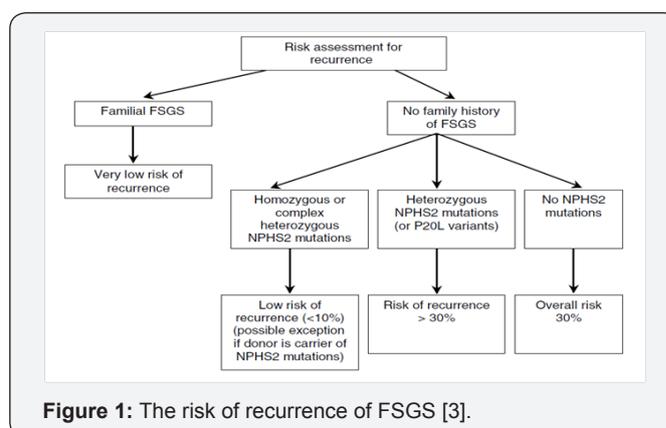
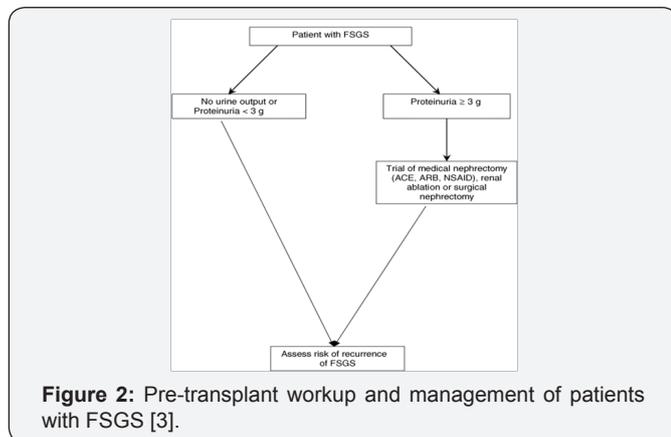


Figure 1: The risk of recurrence of FSGS [3].

The possibility of recurrent FSGS post renal transplantation is about 30% [1,4]. Vincenti et al. [3] have simplified the risk of recurrence in the following algorithm (Figure 1) [3]. Risk factors for recurrence include childhood onset of the original disease,

an aggressive course of rapid progression to ESRD (less than three years), collapsing variant of FSGS, podocin gene mutation and recurrence of FSGS in a previous allograft [1,3]. A paediatric study has addressed initial steroid sensitivity of the primary FSGS as a predictor of recurrence post transplantation (92.9%), in comparison to 30.2% recurrence rate in steroid-resistant cases ($P < 0.001$) [5].

It is advisable that patients with FSGS have minimal proteinuria at the time of transplantation, this will help in early detection of significant proteinuria post transplantation [3]. Thus, patients with significant proteinuria should have a trial of non-steroidal anti-inflammatory drugs (NSAIDs), with or without an angiotensin-converting enzyme inhibitor (ACEI) or angiotensin II receptor blocker (ARB) as illustrated in Figure 2. These drugs may induce what is called medical nephrectomy associated with the shutdown of the remaining kidney nephrons. Moreover, in case this strategy failed, and significant proteinuria persists, nephrectomy may be considered [3].



Early diagnosis of FSGS recurrence will allow early initiation of therapy, which may reverse proteinuria and preserve kidney function [4,6]. However, Recurrent FSGS will result in allograft loss in more than 50% of cases [4,7].

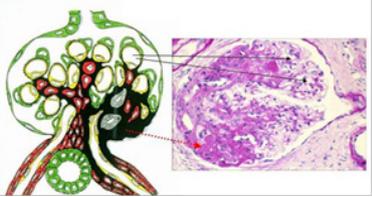
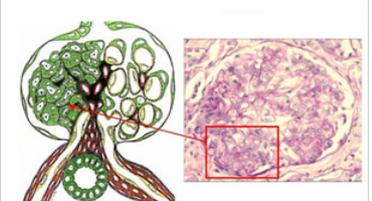
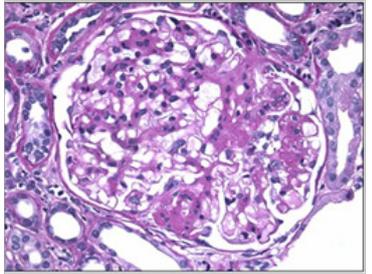
Vincenti et al. [3], suggested a protocol for screening and early detection of recurrence of FSGS. They advised to perform daily protein/creatinine ratio (P/C ratio) in spot urine sample starting from first day post transplantation and throughout the hospital stay, then weekly for one month and monthly after that till the end of the first year [3]. If the ratio is ≥ 0.5 , they advised collecting 24 hours urine sample for quantification of proteinuria. If proteinuria is more than 2g/24 h, then a renal biopsy is indicated [3].

Prediction of FSGS recurrence is of great importance, as it will help in the early implementation of the therapeutic intervention. The prompt and timely initiation of therapy will result in improved patient and graft survival, reduce the cost of late intervention (including hospitalisation and the need for dialysis) and maximise the use of the available transplant organs [8]. Delville et al. [8], had identified a panel of seven antibodies (CD40, PTPRO, CGB5, FAS, P2RY11, SNRPB2, and APOL2). These antibodies if present in the serum of the recipient pre-transplant, it can predict post-transplant FSGS recurrence with 92% accuracy [8].

The pretransplant high titre of anti-CD40 antibody alone is significantly correlated with post-transplantation FSGS recurrence with about 78% accuracy [8]. Furthermore, the injection of purified human anti-CD40 antibodies, isolated from the serum of recipients with recurrent FSGS into experimental mice has led to the development of proteinuria. The proteinuria induced by these antibodies was considerably potentiated when injected with the soluble urokinase receptor (suPAR). On the other hand, injection of a CD40-blocking antibody was documented to significantly reduce the proteinuria induced by anti-CD40 antibodies in the affected mice ($ACR 380.2 \pm 94.3$ before versus 67.8 ± 2.8 after blocking; $P = 0.028$) [8]. The investigators concluded that further evaluation of this antibody panel might provide a tool for pre-emptive therapeutic intervention via plasmapheresis, immunoabsorption, B lymphocyte suppression or disruption of the CD40axis by a CD40-blocking antibody [8].

Table 1: FSGS variants identified by Columbia classification [10,11].

Name of the Variant	Descriptive Histology	
Collapsing variant	At least one glomerulus with segmental or global collapse and overlying podocyte hypertrophy and hyperplasia.	
Tip variant	At least one segmental lesion involving the tip domain (outer 25% of tuft next to origin of proximal tubule).	
	The tubular pole must be identified in the defining lesion.	
	The lesion must have either an adhesion or confluence of podocytes with parietal or tubular cells at the tubular lumen or neck.	
	The tip lesion may be cellular or sclerosing.	

<p>Perihilar variant</p>	<p>At least one glomerulus with perihilar hyalinosis, with or without sclerosis.</p> <p>More than 50% of glomeruli with segmental lesions must have perihilar sclerosis and/or hyalinosis.</p>	 <p>The image shows a schematic of a kidney with a red circle highlighting a glomerulus. To the right is a corresponding histological section showing a glomerulus with a prominent pink-stained perihilar region, indicating hyaline sclerosis.</p>
<p>Cellular variant</p>	<p>At least one glomerulus with segmental endocapillary hypercellularity occluding the lumen, with or without foam cells and karyorrhexis.</p>	 <p>The image shows a schematic of a kidney with a red circle highlighting a glomerulus. To the right is a corresponding histological section showing a glomerulus with a red box highlighting an area of hypercellularity within the capillary lumen.</p>
<p>Not otherwise specified (NOS) variant</p>	<p>At least one glomerulus with a segmental increase in matrix obliterating the capillary lumen.</p> <p>There may be segmental glomerular capillary wall collapse without overlying podocyte hyperplasia.</p>	 <p>The image is a histological section showing a glomerulus with a large area of pink-stained matrix that has significantly narrowed the capillary lumen.</p>

Other investigators tried to correlate the risk of recurrence with the initial histopathological findings in the native kidney biopsy [9]. The Columbia classification was introduced in 2004, and it classifies FSGS into five distinct variants based on their histological appearance in renal biopsy as illustrated in Table 1 [10].

Despite the good correlation between the Columbia classification and the clinical course of FSGS in the native kidneys, it was of no help in the prediction of recurrence post transplantation. Moreover, the histopathological findings in a case of recurrence were different from those of the native kidney, and this difference exists in the first, second and even in the third allograft [9].

The treatment of FSGS recurrence is uncertain with no available international guidelines; few studies reported beneficial results of plasmapheresis (ranging from 10-23 sessions), with or without rituximab [12]. Moreover, some centres combined plasmapheresis with cyclophosphamide (while the anti-metabolite immune suppression transiently discontinued) [13,14]. Both strategies showed promising results with complete or partial remission of FSGS. A reasonable approach is to start with ACEIs or ARBs in the case of persistence of significant proteinuria, then proceed to one of the above-listed approaches depending on the patient medical condition, the potential side effects of each intervention and on the local resources.

Conclusion

The clinical course of primary FSGS can predict the recurrence in the renal allograft, while the variant of the primary disease

may not have the same predictive ability. Identification of new antibodies which may be contributing to the disease recurrence and subsequently directing the treatment is a promising approach in the management of this graft-threatening condition.

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