



Effect of Various Factors in Blood Parameters of Laying Hens in Cages Indoor Grown in Afyonkarahisar



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Abstract

This study was carried out in five hundred Hy Line breed chickens at the 60 days age breeding in Afyonkarahisar Province. This study was done during the winter season when the outside temperature was 13 °C, while the average indoor temperature of the shelters was 23°C. Clinical, hematological and blood biochemical parameters were measured in chickens that constituted the study material. Clinically; body temperature (T), respiration (R) and heart rates (P) were measured. In the hematological examinations; formula leukocyte counts were performed with erythrocyte (RBC), total leukocyte (WBC), hemoglobin (HB), hematocrit (HCT), mean corpuscular volume (MCV), mean erythrocyte hemoglobin (MCH) and mean hemoglobin concentration (MCHC). In the blood biochemical examinations; serum aspartate aminotransferase (AST), serum gamma-glutamyltransferase (GGT), ornithine carbamoyltransferase (OCT), serum lactate dehydrogenase (LDH), serum sorbitol dehydrogenase (SDH) and creatine phosphokinase (CPK), total protein (TP), Albumin (ALB), Glucose (GLU), Total and Indirect Bilirubin (TB and IB), Total Cholesterol (TCOL), High Density Lipoprotein (HDL) and Low Density Lipoprotein (LDL) levels were determined. At the end of the study; it has been observed that all of the measured parameters differ from the results obtained from earlier studies, although those within the normal limits. The probable cause of this variability was linked to changes in race, nutritional and climatic conditions. The results obtained from the study were the first to examined clinical, hematological and blood biochemical parameters in chickens breeding in closed cages in Afyonkarahisar Province, and then to reference the field studies of scientific studies and practicing veterinarians to study about this subject.

Keywords: Chicken; Clinical; Hematological; Blood biochemistry

Introduction

The world population will increase by 33% by 2050, which is predicted to lead to a 70% increase in total food production [1], leading to an increase in poultry meat and egg demand. Chicken meat and protein derived from eggs is the cheapest source of meat for human consumption, the United Nations Food and Agriculture Organization (FAO), behind the pig of the poultry world population is ranked as the second largest source of protein [2]. However, the commercial poultry industry, infection that can affect productivity negatively, is faced with stress factors such as feed variations and climate change [3,4]. Indeed, the poultry growth performance is not only inherited, but is also greatly influenced by the surrounding [5]. Stress directly affects the physiology and well-being of poultry and results in a lower rate of return [6,7].

Among the most common stress factors in cage-type poultry farming are; temperature, humidity, lack of light, hosting crowd, ventilation, noise and fear, infection, vaccination and transport are to count [8-10]. Many physiological and metabolic activities

are formed in response to stress events, which may result in pathological changes in hematological and serum biochemical parameters [11].

In this study; it was aimed to present the first case of important stress, hematological and biochemical parameters for internal diseases in chickens grown in modern facilities.

Materials and Methods

Animal material

The material of this work was the 500 Hy-Line raki 60-day-old egg chickens grown with closed cage system in Afyonkarahisar Province. The study was carried out in winter and the average temperature of the plants was measured at 23°C. It was determined that the plant was kept at these temperatures continuously during the winter season and the same rations were given to the animals and they were grown under the same conditions in the same scaled cages, the calculation continued with the calculation of the arithmetic mean (AO) of the parameters being measured.

This study has been carried out with the reference number of AKUHADYEK 344-16 under the ethics rules of Afyon Kocatepe University Animal Experiments Ethics Committee and it was supported by the reference number of 17.SGBIL.16 and Afyon Kocatepe University Scientific Research Projects Coordination Unit (BAPK).

Methods

Clinical experiments: Body temperature, nostril observation, auskultation, respiration and heart rate were measured by cloacal route and animals were recorded to be evaluated.

Hematological measurements: In animals, blood was drawn into EDTA blood tubes via brachial ven so that it would not exceed 1% of live weight, blood samples were sent to the laboratory within the same day and as soon as possible. In blood samples taken for hematological examination; (RBC), total leukocyte (WBC), hematocrit (HCT), hemoglobin (HB), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MHC), mean corpuscular hemoglobin concentration (MCHC), lymphocyte (LENF), neutrophil Hematological examinations such as NOTR, eosinophil (EOS), monocyte (MON) and basophil (BAZ) were measured using commercial test kits with a Chemray Brand blood count device.

Serum biochemical measurements: For this purpose, sera of the blood samples taken at the site were taken out and the extracted sera were stored at +4°C by taking gods if not immediately measured. In blood biochemical examinations; Serum aspartate aminotransferase (AST), serum gamma glutamyltransferase (GGT), ornithyl carbonyl transferase (OCT), serum lactate dehydrogenase (LDH), serum sorbitol dehydrogenase (SDH) (spectrophotometrically measured at 340nm wavelength on a Chemwell Mark device) (TPL), albumin (ALB), glucose (GLU), total and indirect bilirubin (TB and IB), total cholesterol (TCOL), high density lipoprotein (HDL) and low density lipoprotein) Were determined using commercial kits in the Chemwell Brand autoanalyser.

Statistical analyses

The existence of similar stress factors in all of the animals that make up the material of the study, the lack of significant differences in terms of the stress parameters among the birds, and the similarity of the projects, ration, altitude, (AO) (Yildiz et al. 2002), the results of some studies considered nominal in terms of the previously mentioned parameters are not available

Table 3: Hayvanların Kan Biyokimyasal Analizleri.

Values	TP (g/dl)	ALB (g/dl)	GLU (mg/dL)	TB (nmol/L)	IB (nmol/L)	TCOL (mg/dL)	HDL (mg/dL)	LDL (mg/dL)	AST (IU/L)	GGT (IU/L)	LDH (IU/L)	CK (IU/L)	SDH (IU/L)
AO	3.96	1.74	243	0.005	-	138.5	27.71	74.345	251.25	18.7	1147.35	2651.35	0.026
ED-EY	2.62-5.30	1.14-2.34	126-360	0-0.01	-	78-199	21.82-33.6	62.39-86.3	164.5-338	10-27.4	960.5-1334.2	1870.4-3432.3	0.019-0.033

levels were found to be appropriate.

Results

In our present study, the temperature measured during the inspection and inspection at the enterprises was determined as 23°C, but there was no possibility to measure the humidity during this operation. The facilities are built according to European standards and this includes stress, such as the presence of sufficient light and ventilation requirements, the fact that indoor climate conditions are prevalent, animals are raised in crowded places, the high temperature due to the crowd in special areas, The presence of factors has been identified. As a matter of fact, it was observed that around 30 animals were housed in a section of 20 animals during the study.

Clinical findings

In our current study, the AOs of body temperature, respiration, and heart rates and the lowest and highest values of the animals in the material forming are shown in Table 1.

Table 1: The lowest and highest levels of body temperature, heart and respiratory freaks in animals were obtained with AOs.

Values	T (°C)	P (freq/min)	R (freq/min)
AO	40.05	274	44.5
ED-EY	39.4-40.7	230-318	34-55

Hematological findings

The AOs of the hematological parameters measured for the animals forming the material and their lowest and highest levels are shown in Table 2.

Table 2: Measured hematological parameters.

Values	T (°C)	P (freq/min)	R (freq/min)
AO	40.05	274	44.5
ED-EY	39.4-40.7	230-318	34-55

Values	LENF (%)	NOTR (%)	EOS (%)	MON (%)	BAS (%)
AO	57.5	27	4.5	7	2
ED-EY	42-73	15-39	7-Feb	11-Mar	3-Jan

AO: Arithmetic mean, ED-EY: Low-highest values

Serum biochemical findings

Blood biochemical analysis endpoint AOs measured in this study and the lowest and highest measured values are shown in Table 3.

Discussion

Stress is the name given to all of the biochemical, physiological and behavioral changes that occur in the body in order to restore the distorted hemostatic balances of animals exposed to the factors [12]. It has been reported in studies conducted in animals exposed to hunger and transport stress that the sympatho-adrenal system is stimulated in stress cases, which causes an increase in body temperature, respiration and cardiac frequency (Srikandakumar and Johnson) [13]. It has also been pointed out that the increase in body temperature in homothermic animals in hot and humid conditions is expected [14,15]. In this study, we found that the mean body temperature, respiration, and heart rate were within normal limits, but the averages were higher than those reported by the investigators [16]. As a matter of fact, in hot environmental conditions chickens open their mouths and increase respiration rates and respiration rates. The reason for this is to reduce body temperature by throwing water vapor from the lungs. As the ambient temperature increases, the breathing frequency increases and the need for metabolizable energy increases [17-19] to keep the animal breathing in order to reduce body temperature.

In the case of increased stresses such as temperature and humidity, the immune system is suppressed, and as a result, resistance to the disease has been reduced [20,21]. In this study, we found significant changes in the haematological tabulation of chickens grown under relatively stress conditions, an increase in the number of NOTRs compared to the above-mentioned studies, and a lower level in the LENF number averages. Similar findings are consistent with studies [22-24] that show that poultry treated with CS or ACTH has a reduced number of circulating lymphocytes. This decrease is due to the increase in the proportion of circulating heterophiles to lymphocytes, which is considered to be one of the most common stress symptoms in birds [25], ie the increase in heterophil/ lymphocyte ratio. This decrease in lymphocyte counts is probably due to lymphoid tissue tension due to long duration of stressors [26,27]. As a matter of fact, [28] stated that lymphocytes were consumed in germinal centers after ACTH or CS injections and that lymphocyte production was inhibited by lymphoid tissue atrophy (Virden and Kidd, 2009). Studies of heat stress on haematological parameters and macrophage activity have also shown that macrophage activity decreases due to temperature and heat stress up to 36°C, which reduces intestinal injury, leading to an increase in the population of pathogenic bacteria along the intestinal epithelium and also to infectious stress in poultry (Quinteiro-Filho et al. 2012; Verbrugge et al. 2012).

Along with changes in hematological parameters during stress, there are also significant changes in blood composition. As a matter of fact, catabolism is promoted in order to meet the nutritional requirements for the synthesis of immune effector molecules while weakening body proteins and fat anabolism under immune stress [29,30]; If corticosteroids occur at high

levels in the circulation; glucose and mineral metabolism, resulting in structural disorders such as cardiovascular disorders and hypercholesterolemia [31-33].

In the literature reviews we made; it has been determined that the results obtained in the studies investigating the blood biochemical parameters of the same breed in similar conditions and the values obtained in our study are different. Schaal et al. [34] found GLU levels to be 224, while our study found 243mg/dL. Again, different results of blood biochemical measurements were obtained from other studies with the same parameters. Gynesis et al. [35] conducted three phenotypes of the same breed of chicken in all three phenotypes; They found TP <5 (g / L), ALB (g / L) <2, HDL (mmol / L) <3, CPK <3000 (U / L) and LDH <2500 (U / L). Khawaja et al. (2012) reported different values for the results of our study as GLU 221 (mg/dL), cholesterol 138.75 (mg/dL), TP 05.10 (mg/dL).

Exposure of catecholamines to epinephrine and norepinephrine is increased when the strase is exposed. Especially, catecholamines play the most important role in altering epinephrine metabolism [31,36] Sabban and Kvetnansky, 2001. Epinephrine binds to β -adrenergic receptors on the cell membrane and results in an increase in the activation of certain enzymes, primarily the activation of protein kinases that activate glycogenolysis and gluconeogenesis Olanrewaju et al. 2006; [37]. In stress conditions, norepinephrine is responsible for providing energy from the fatty tissue for skeletal muscles [38,39]. In addition to GLU levels, some enzymes such as AST, GGT, and CHOL levels increase in tissues and tissues, mainly in muscle tissue, as a result of catabolic activities in tissues and organs [40-42]. In this study, similar to the ones reported by the researchers, it is proved that the levels of AST, GGT are close to the upper limits of the normal limits.

When compared to the studies we have conducted, the detection of very low levels of TB and no detection of IB levels is consistent with the report that chickens have insufficient production of bilirubin [43,44].

The obtained data also suggests that it is an original work at the same time that it is the quality that will form the reference for the Hy Line chickens raised in the region. We believe that it is essential for the poultry sector of our country and our country to investigate stress events affecting fertility in more detail [45-50].

References

1. (2015) UNO - United Nations Organisation. Department of Economic and Social Affairs.
2. Lopez JC (2006) The Effect of Environmental Stressors on The Immune Response To Avian Infectious Bronchitis Virus. Thesis. Lincoln University, USA, pp. 1-161.
3. Shini S, Kaiser P, Shini A, Bryden WL (2008) Biological response of chickens (*Gallus gallus domesticus*) induced by corticosterone and a bacterial endotoxin. *Comp Biochem Physiol B Biochem Mol Biol* 149(2): 324-333.

4. Mengesha M (2004) Climate change and the preference of rearing poultry for the demands of protein foods. *Asian Journal of Poultry Science* 5(4): 135-143.
5. Babinszky L, Halas V, Versteegen MW (2011) Impacts of climate change on animal production and quality of animal food products. In: Kheradmand H (Ed.), *Climate change socioeconomic effects*. InTech, Rijeka, Hungary, pp. 165-190.
6. Barnett J, Hemsworth P (2003) Science and its application in assessing the welfare of laying hens in the egg industry. *Aust Vet J* 81(10): 615-624.
7. Osti R, Bhattarai D, Zhou D (2017) Climatic Variation: Effects on Stress Levels, Feed Intake, and Bodyweight of Broilers. *Brazilian Journal of Poultry Science* 19(3): 489-496.
8. Yahav S, Goldfeld S, Plavnik I, Hurwitz S (1995) Physiological responses of chickens and turkeys to relative humidity during exposure to high ambient temperature. *J Therm Biol* 20(3): 245-253.
9. Zulkifli I, Che Norma MT, Chong CH, Loh TC (2000) Heterophil to lymphocyte ratio and tonic immobility reactions to preslaughter handling in broiler chickens treated with ascorbic acid. *Poul. Sci* 79(3): 402-406.
10. Wei FX, Hu XF, Xu B, Zhang MH, Li SY, et al. (2015). Ammonia concentration and relative humidity in poultry houses affect the immune response of broilers. *Genet Mol Res* 14(2): 3160-3169.
11. Chikumba N, Swatson H, Chimonyo M (2013). Haematological and serum biochemical responses of chickens to hydric stress. *Animal* 7(9): 1517-1522.
12. Konca Y, Yazgan O (2002) Yumurta Tavuklarında Sıcaklık Stresi ve Vitamin C. *Hayvansal Üretim* 43(2): 16-25.
13. Ait-Boulahsen A, Garlich JD, Edens FW (1989) Effect of fasting and acute heat stress on body temperature, blood acid-base and electrolyte status in chickens. *Comp Biochem Physiol A Comp Physiol* 94(4): 683-687.
14. Darcan N (2000) A study on adaptation mechanism of crossbred goat types in Çukurova subtropical climate conditions. Çukurova Univ, Institute of Science, Adana, Turkey.
15. Star L, Kemp B, Van Den Anker I, Parmenter HK (2008) Effect of single or combined climatic and hygienic stress in four layer lines: 1. Performance. *Poultry Science* 87(6): 1022-1030.
16. Rozenboim I, Biran I, Uni Z, Robinson B, Halevy O (1999) The effect of monochromatic light on broiler growth and development. *Poult Sci* 78(1): 135-138.
17. Ayo JO, Oladele SB, Fayomi A, Jumbo SD, Hambolu JO (1998) Body temperature, respiration and heart rate in the Red Sokoto goat during the harmattan season. *Bull Anim Hlth Prod in Afr* 46: 161-166.
18. Diarra SS, Tabuaciri P (2014) Feeding Management of Poultry in High Environmental Temperatures. *International Journal of Poultry Science* 13(11): 657-661.
19. Franco-Jimenez DJ, Beck MM (2007) Physiological changes to transient exposure to heat stress observed in laying hens. *Poult Sci* 86(3): 538-544.
20. Daynes RA, Araneo BA (1989) Contrasting effects of glucocorticoids on the capacity of T cells to produce the growth factors interleukin 2 and interleukin 4. *Eur J Immunol* 19(12): 2319-2326.
21. Blecha F (2000) Immune system response to stress. In: Moberg GP, Mench JA (Eds.), *The Biology of Animal Stress*. CABI Publishing, UK, pp. 111-121.
22. Garren HW, Shafner CS (1956). How the period of exposure to different stress stimuli affects the endocrine and lymphatic gland weights of young chickens. *Poult Sci* 35(2): 266-272.
23. Meyer RK, Aspinall RL, Graetzer MA, Wolfe HR (1964) Effect of corticosterone on the skin homograft reaction and on the precipitin and hemagglutinin production in thymectomized and bursectomized chickens. *J Immunol* 92: 446-451.
24. Griffin JF (1989) Stress and immunity: A unifying concept. *Vet Immunol Immunopathol* 20(3): 263-312.
25. Segel HS (1995) Stress, strains, and resistance. *Br Poult Sci* 36(1): 3-22.
26. Huff GR, Huff WE, Balog JM, Rath NC, Anthony NB, et al. (2005) Stress response differences and disease susceptibility reflected by heterophil to lymphocyte ratio in Turkeys selected for increased body weight. *Poult Sci* 84(5): 709-717.
27. Dunkley CS, McReynolds JL, Dunkley KD, Njongmeta LN, Berghman LR, et al. (2007) Molting in Salmonella-Enteritidis-challenged laying hens fed alfalfa crumbles. IV. Immune and stress protein response. *Poultry Science* 86(12): 2502-2508.
28. Glick B (1967) Antibody and gland studies in cortisone and ACTH-injected birds. *J Immunol* 98(5): 1076-1084.
29. Piquer FJ, Sell JL, Sotosalanova ME, Vilaseca L, Palo PE, Turner K (1995) Effects of early immune stress and changes in dietary metabolizable energy on the development of newly hatched turkeys. 1. Growth and nutrient utilization. *Poultry Science* 74(6): 983-997.
30. Lin H, Decuypere E, Buyse J (2004) Oxidative stress induced by corticosterone administration in broiler chickens (*Gallus gallus domesticus*) 1. Chronic exposure. *Comp Biochem Physiol B Biochem Mol Biol* 139(4): 737-744.
31. Carter AM, Petersen YM, Towstoles M, Andreassen D, Jensen BL (2002) Adrenocorticotrophic hormone (ACTH) stimulation of sheep fetal adrenal cortex can occur without increased expression of ACTH receptor (ACTH-R) mRNA. *Reprod Fertil Dev* 14(1-2): 1-6.
32. Virden WS, Lilburn MS, Thaxton JP, Corzo A, Hoehler D, et al. (2007) The effect of corticosterone-induced stress on amino acid digestibility in Ross broilers. *Poult Sci* 86(2): 338-342.
33. Olanrewaju HA, Miller WW, Maslin WR, Collier SD, Purswell JL, et al. (2016) Effects of light sources and intensity on broilers grown to heavy weights. Part 1: Growth performance, carcass characteristics, and welfare indices. *Poult Sci* 95(4): 727-735.
34. Schaal TP, Arango J, Wolc A, Brady JV, Fulton JE, et al. (2016) Commercial Hy-Line W-36 pullet and laying hen venous blood gas and chemistry profiles utilizing the portable i-STAT®1 analyzer. *Poult Sci* 95(2): 466-471.
35. Gyenis J, Suto Z, Romvari R, Horn P (2006) Tracking the development of serum biochemical parameters in two laying hen strains -a comparative study. *Arch Tierz Dummerstorf* 6(2): 593-606.
36. Solemani AF, Zulkifli I, Omar AR, Raha AR (2011) Physiological responses of 3 chicken breeds to acute heat stress. *Poult Sci* 90(7): 1435-1440.
37. Kim JA (2008) Mechanisms underlying beneficial health effects of tea catechins to improve insulin resistance and endothelial dysfunction. *Endocr Metab Immune Disord Drug Targets* 8(2): 82-88.
38. Castillo M, Amalik F, Linares A, García-Peregrín E (1999) Dietary fish oil reduces cholesterol and arachidonic acid levels in chick plasma and very low density lipoprotein. *Mol Cell Biochem* 200(1-2): 59-67.
39. Huang J, Zhang Y, Zhou Y, Zhang Z, Xie Z, et al. (2013) Green tea polyphenols alleviate obesity in broiler chickens through the regulation of lipid-metabolism-related genes and transcription factor expression. *J Agr Food Chem* 61(36): 8565-8572.
40. Ramage-Healey L, Romero LM (2001) Corticosterone and insulin interact to regulate glucose and triglyceride levels during stress in birds. *Am J Physiol Regul Integr Comp Physiol* 281(3): 994-1003.
41. Ashwell CM, McMurtry JP (2003) Hypoglycemia and reduced feed intake in broiler chickens treated with metformin. *Poult Sci* 82(1): 106-110.

42. Altan O, Altan A, Cabuk M, Bayraktar H (2000) Effects of heat stress on some blood parameters in broilers. *Turk J Vet Anim Sci* 24: 145-148.
43. Bromidge ES, Wells JW, Wight PA (1985) Elevated bile acids in the plasma of laying hens fed rapeseed meal. *Res Vet Sci* 39(3): 378-382.
44. Khawaja T, Khan SH, Mukhtar N, Ali Ma, Ahmed T, et al. (2012) Comparative study of growth performance, egg production, egg characteristics and haemato-biochemical parameters of Desi, Fayoumi and Rhode Island Red chicken. *Journal of Applied Animal Research*, 40(4): 273-283.
45. Mendl M (1999) Performing under pressure stress and cognitive function. *Applied Anim Behav Sci* 65(3): 221-244.
46. Sundrum A (2001) Organic livestock farming: A critical review. *Livestock Prod Sci* 67(3): 207-215.
47. Sekeroglu A, Demir E, Sarica M, Ulutas Z (2009) Effects of Housing Systems on Growth Performance, Blood Plasma Constituents and Meat Fatty Acids in Broiler Chickens. *Pak J Biol Sci* 12(8): 631-636.
48. Curi TMRC, Conti D, Vercellinor A, Massari JM, Moura DJ, et al. (2017). Positioning of sensors for control of ventilation systems in broiler houses: a case study. *Scientia Agricola* 74(2): 101-109.
49. Barnett JL, Hemsworth PH, Newman EA (1992) Fear of humans and its relationships with productivity in laying hens at commercial farms. *Br Poult Sci* 33(4): 699-710.
50. Yarsan E, Gülec M (2003) Kanatlarda stres, vitamin ve mineral uygulamaları. *Türk Veteriner Hekimleri Birliği Dergisi*, p. 55-63.



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