Cochlear Endoscopy in Cochlear Implantation of a X-Linked Stapes Gusher Syndrome

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Abstract

A 12-year-old boy with a five-year history of bilateral sensorineural hearing loss and X-linked stapes gusher syndrome developed progressive left-sided hearing loss. The pre-operative computed tomography of the temporal bone showed a bulbous internal auditory canal with a dysplastic cochlea and no apparent modiolus. A 1.3mm salivary endoscope was placed at the cochlear entrance to assess the intracochlear anatomy. This revealed membranous structures of the cochlea without direct communication to the internal auditory canal. We advocate for the use of cochlear endoscopy to better delineate inner ear anatomy, which will influence the implant selection and potentially hearing outcomes in patients.

Keywords: Cochlear endoscopy; Otoendoscopy; X-linked stapes gusher syndrome; Cochlear implant; Perilymphatic gusher.

Introduction

X-linked stapes gusher syndrome (otherwise known as X-linked deafness type 3 or Nance deafness) is a rare form of sensorineural hearing loss (SNHL) syndrome. Inherited in a sex-linked recessive manner, it is believed to be the consequence of a loss-of-function mutation on the X-chromosome in the POU3F4 gene at the DFN3 locus [1,2]. Males tend to present greater phenotypic severity than females, who present less frequently [3-5].

Affected patients have an abnormal configuration of the lamina cribrosa and internal auditory canal (IAC) [1,3,4,6,7]. This malformation leads to increased perilymphatic pressure and to stapes’ footplate fixation, giving rise to conductive hearing loss and progressive cochlear nerve incompetence. This is relevant for surgeons, as it results in an increased risk of perilymph gusher with surgical manipulation [1,3,4,6,7]. This may lead to other complications, such as otorrhea, rhinorrhea, and recurrent meningitis [8]. Meningitis complicating cochlear implantation (CI), occurs at a higher rate in patients with inner ear (IE) abnormalities, including X-linked gusher syndrome [3,9].

Traditional approaches to imaging for CI utilize computed tomography of the temporal bone (CT TB) and/or magnetic resonance imaging. We report a case whereby intra-operative otoendoscopic visualization allowed for real time visualization of the IE anatomy, which allowed us to optimize our electrode choice for CI. In this case, such visualization was valuable, as it indicated the presence of the membranous portions of the IE to evaluate if the modiolus was present, which we believe implied that a directional electrode was the most appropriate choice.

Case Report

A 12-year-old boy with a five year history of bilateral SNHL and X-linked stapes gusher syndrome presented with progressively worsening left-sided hearing loss. He was initially performing well with bilateral traditional hearing aids but developed progressive mixed loss and worsening performance in his left ear. His preoperative audiogram can be seen in Figure 1. He was sent for CI candidacy assessment and was deemed to be suitable.
Figure 1: Pre-operative audiogram with speech testing demonstrating mixed loss and worsening performance in the left ear.

Figure 2: Computer Tomography of the Temporal Bone that shows a dysplastic cochlea with no apparent modiolus.

The pre-operative computed tomography (CT) of the temporal bone was assessed. This showed a bulbous IAC with a dysplastic cochlea and no apparent modiolus (Figure 2). A CI24RE (ST) implant was initially selected by the implant team due to the uncertainty regarding the location of the spiral ganglion cells.

A 1mm cochleostomy was made anteroinferior to the round window (RW). Upon entering the cochlea, a moderate perilymphatic gusher was encountered. A 1.3mm rigid salivary endoscope (Karl Storz, Germany) with a 3-chip full HD camera head (Image1 S H3-Z) was approached transtympanically and was placed at the edge of the 1mm cochleostomy to assess the intracochlear anatomy.

To avoid injury to the inner ear, the CSF was not suctioned. The same view was not achieved with the microscope as the endoscope provided a more magnified and higher definition view of the modiolus. There was evidence of membranous structures of the cochlea without direct communication to the IAC. Full insertion was achieved on the first attempt without any difficulty and the gusher was controlled with packing of periosteal tissue at the cochleostomy site. An intra-operative plain X-ray confirmed the electrode placement.

Discussion

When the cochlear modiolus and osseous spiral lamina are deficient, the absence of a bony septum creates a common space seen on the imaging study [10] (Figure 2); there is an abnormal communication between the IAC and the vestibule responsible for the perilymphatic gusher. Patients with IE anomalies may have both atypical positions of their spiral ganglion cells (SGC) and have a higher likelihood of having fewer spiral ganglion cells [11]. This has functional implications because at least 10,000 functional SGNs are necessary for effective speech discrimination.
abnormal. In patients that have an absent modiolus, a circumferentially stimulating electrode may be preferred over a full-banded electrode, which may risk adverse facial nerve stimulation [3]. One explanation for post-operative facial nerve stimulation in children with IE abnormalities is the close vicinity of the electrode to the nerve [14]. Therefore, to avoid injury, the proper choice in electrode should be made.

**Conclusion**

While CT temporal bone has served as the conventional approach to assessing the anatomy of the IE, the endoscope offers better resolution of the modiolus than the CT temporal bone, as the CT indicated that there was no modiolus. We advocate the use of intracochlear endoscopy in selected cases as it offers a better resolution than even high-resolution CT. With the potential to change electrode choices in CI, the customization of electrode choice based on the presence of membranous IE anatomy may change the hearing outcome of the patients with anomalous IE anatomy and patients with uncertain location of spiral ganglion cells.

**References**


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