



Crustacean Endocrinology: Intriguing Towards Quality Protein Production



Ramachandra Reddy P^{1,2*} and Kishori B³

¹Department of Biochemistry, Yogi Vemana University, India

²Department of Food Science, Pennsylvania State University, USA

³Department of Biotechnology, Sri Padmavathi Mahila Viswavidyalayam, India

Submission: February 20, 2017; Published: March 07, 2017

*Corresponding author: Ramachandra Reddy P, Department of Biochemistry, Yogi Vemana University, Vemanapuram, Kadapa-516 003, Andhra Pradesh, India, Department of Food Science, Pennsylvania State University, USA, Tel: +91-08562-225425, Fax: +91-08562-225419; Email: prcrbio@yogivemanauniversity.ac.in

Abstract

Present World in dearth of nutritious food, especially proteinaceous food forever growing human population. Alternatives are in search in producing more high quality protein food in a short time with available resources. The crustacean aquaculture is one of the important agricultural fields in producing high quality proteinaceous food, but it is having its own limitations to produce more amount of protein food. However, manipulation of endocrine system is one of the best ways to increase crustacean protein food. This article reviews the possible ways of crustacean endocrine manipulations to promote the growth to increase protein productivity in crustacean aquaculture industry.

Introduction

Malnutrition is the major problem in the developed, developing and third World countries where the people's diet does lacking adequate calories and protein for maintenance and growth. Supplementation of nutritious food to everyone in the World is a big task ahead, to achieve this more amount of quality food, especially protein should be produced with the available resources. Agriculture is the main source of producing protein of which aquaculture plays an important role in producing quality protein. Both finfish and shellfish come under aquaculture. Finfish culture is much better in producing protein than shellfish culture. In shellfish, crustaceans are most important animals to produce quality protein which has high value Worldwide.

Many studies are under taken to improve the crustacean protein in the past. One of the classical method followed to increase the growth in crustaceans is by eyestalk ablation unilaterally or bilaterally [1-6]. Induction of precocious molting has also been well documented in the spider crab, *Libinia emarginata* [7], in the freshwater crab *Oziothelphusa senex senex* [8] and the freshwater prawn, *M. rosenbergii* after eyestalk ablation [9]. Though eyestalk ablation induces the growth and reproduction in cultured species, but has its own limitations like loss of hemolymph by cautery, mortality in the ablated animals and hormonal imbalance since an array of hormones synthesized and secreted from eyestalk are removed [10-12].

Table 1: CHH family peptides and their overlapping biological activities.

S. No.	Chh-Family Peptide	Specific Activity	Overlapping Activity	References
1	CHH	Glucose metabolism and Osmoregulation	Molting and Reproduction	Jeon et al. [42], Webster et al. [57]
2	MIH	Molting	Reproduction and Glucose metabolism	Reddy et al. [3], Zmora et al. [58], Luo et al. [48]
3	MOIH	Molting and Reproduction	Glucose metabolism	Wainwright et al. [25], Lu et al. [47], Nagaraju et al. [50], Reddy and Reddy [2,3]
4	GIH/VIH	Reproduction	Glucose metabolism and Molting	De Kleijn and Van Herp [41], Kang et al. [43]
5	RPCH	Exoskeleton pigmentation	Ovarian maturation and molting	Kulkarni et al. [30], Sarojini et al. [19], Swetha et al. [18]

Eyestalk in crustaceans is a reserve for many hormones regulates several physiological functions comprising growth and reproduction. An array of hormones released from crustacean eyestalk includes crustacean hyperglycemic hormone (CHH), molt-inhibiting hormone (MIH), mandibular organ-inhibiting

hormone (MOIH) and gonad/vitellogenesis-inhibiting hormone (VIH) besides other hormones and neurotransmitters (see reviews [2,13-16]. These eyestalk neuropeptides are collectively called as CHH-family peptides since they share the functional similarity and show multiple biological functions (Table 1).

Table 2: The synthesizing, storage and releasing site, nature and functionality of various crustacean endocrine hormones involved in the regulation of growth.

S.No.	Hormone	Synthesizing site	Storage site	Releasing site	Nature	Function	References
1	CHH	Eyestalk ganglion	Sinus gland	Hemolymph	Neuropeptide	Glucose and glycogen metabolism, Osmoregulation, Molting and Reproduction	Keller and Sedlmeier [44], Santhos et al. [53]
2	MIH	Eyestalk ganglion	Sinus gland	Hemolymph	Neuropeptide	Molting, Reproduction and Glucose metabolism	Chang [39] Chang and Mykles [38], Watson et al. [56]
3	MOIH	Eyestalk ganglion	Sinus gland	Hemolymph	Neuropeptide	Molting, Reproduction and Glucose metabolism	Tamone et al. [54], Borst et al. [34], Reddy et al. [4], Nagaraju et al. [50]
4	GIH/VIH	Eyestalk ganglion	Sinus gland	Hemolymph	Neuropeptide	Reproduction and Molting	Quackenbush, [52], Aguilar et al. [32], De Kleijn et al. [41]
5	Ecdysteroids	Y-organ	Y-organ	Hemolymph	Steroid	Molting and Reproduction	Chung, [20] Nakatsuji et al. [51]
6	MF	MO	MO	Hemolymph	Terpenoid	Molting and Reproduction	Nagaraju et al. [51] Suneetha et al. [27] Hemalatha et al. [28]
7	Serotonin	Eyestalk ganglion	Sinus gland	Hemolymph	Hydroxy tryptamine	Neurotransmission and Molting	Sainath and Reddy [29]
8	Leucine-enkephalin	Eyestalk ganglion	Sinus gland	Hemolymph	Tripeptide	Glucose metabolism and molting	Kishori and Reddy [45], Nagabhushanam et al. [19]

Besides eyestalk hormones, the sesquiterpenoid hormone methyl farnesoate (MF), various types of ecdysteroid hormones and many other neurotransmitters show effects on growth (see reviews [14,17,18]. The synthesizing, storage and releasing site, nature and functionality of these molecules are presented in (Table 2). This review emphasizes, an over view of crustacean endocrine hormones and their manipulations towards better growth and quality protein production.

CHH-family neuro peptide hormones

The X-organ sinus gland complex located in the eyestalks synthesizes and secretes CHH-family neuro peptides. The main feature of CHH-family peptides is structural similarity. Due to this they execute multiple functions and shows overlapping biological functions [3]. MIH of these group show inhibitory function on growth and the other peptides also show inhibition as a subsidiary function (Table 1). Manipulation of MIH expression, a way to nullify the inhibitory function of MIH on growth. Recent studies are focused on manipulating this hormone either by removal of eyestalk a classical method bearing several disadvantages and an alternatively by suppressing its activity through RNAi mechanisms [4]. Since most of the CHH-family peptides are having dual biological functions and exhibits inhibitory action on molting of crustaceans (Figure 1), the better way to increase growth is by gene knockout studies. Though the success rate of gene knockout studies is very less, more studies to stabilize these experiments

are essentially done by the researchers.

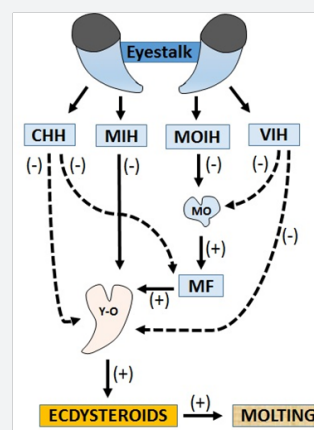


Figure 1: Biological functions of CHH-family neuropeptides. CHH: Crustacean Hyperglycemic Hormone; MIH: Molt-inhibiting Hormone; MOIH: Mandibular Organ-inhibiting Hormone; VIH: Vitellogenesis-inhibiting Hormone; MF: Methyl Farnesoate; MO: Mandibular Organ; Y-O: Y-organ; '+': Induction; '-': Inhibition.

Ecdysteroids

The Y-organ of crustaceans is responsible to produce a group of steroid hormones collectively called as ecdysteroids. The primary function of ecdysteroids in crustaceans is promoting growth [13,14,19]. In another words, increased ecdysteroid levels promotes animal from intermolt to molt (ecdysis) irrespec-

tive of other factors. Since they coordinate molting process they can also be called as molting hormones and the Y-organ as molting gland. These hormones along with another hormone methyl farnesoate promote molting thereby increases the weight of the animal. Many studies are contemplated on ecdysteroid induced growth mostly by injecting these molecules in vivo and in vitro incubation studies [20].

Since ecdysteroid comes under family steroids, instead of injecting these molecules directly into the crustacean for increasing growth, it is better to increase the steroidogenic pathway which in turn increases ecdysterodiogenesis in Y-organs thereby growth of the animal. The Probiotic approach is one of the best ways to increase the ecdysterodiogenesis. Probiotics are combination of food material with live microorganisms that promotes the growth of beneficial microbes in the gut. The most commonly used microbes belong to two bacterial divisions, the *Firmicutes* (e.g. *Bacillus* spp., *Lactobacillus* spp., *Lactococcus* spp., *Carnobacterium* spp. etc.) and the *Gammaproteobacteria* (e.g. *Vibrio* spp., *Pseudomonas* spp., *Shewanella* spp. etc.), while yeasts remain rarely studied [21]. These probiotics changes the gut microbiome of a crustacean with increased digestive capacity and assimilation of digested components into the hemolymph thereby activated metabolism keeps the animal healthy. An active and healthy animal molt faster and attains growth in a short time. Nevertheless, beyond these considerations, especially in Asia Probiotic applications have been a practical and commercial reality in shrimp farming for more than 15 years. In China, more than 100 companies are producing different varieties of probiotics for aquaculture industry [22]. However, in aquatic cultured species the most important factor that needs to be considered is how animals raised and fed which influence the autochthonous and allochthonous microbial communities [23].

Probiotics to produce substrates to initiate ecdysteroidogenesis are appropriate to induce molting in crustaceans. This may be the best way to attain growth in crustaceans, where the lacuna is to proceed further in producing high quality and greater volume crustacean protein.

Methyl farnesoate

The sesquiterpenoid hormone synthesized and secreted from a pair of glands called mandibular organs (MOs) is named as methyl farnesoate (MF). MOs are under the control of eyestalk hormone MOIH, which inhibits the MF synthesis. MF involved in both growth and reproduction and promotes both process [3,4,8,24-28]. The roles of MF on molting were well established in variety of crustaceans and are focused on injecting the MF directly into the animal and studied its effects on growth and reproduction in the laboratory. MF is potential inducer of growth in crustaceans [8]. Since it is not practically possible to inject each animal with MF, the alternative to increase MF levels is by supplementation through the food (Probiotic approach) or by suppressing the action of MOIH by RNAi silencing mechanism.

Both the ways are not yet clearly established in crustaceans and is open for crustacean scientists.

Other molecules

Many other molecules secreted in or coming from the environment are capable to induce growth in crustaceans. The best examples are biogenic amines and opioid peptides. They synthesized in crustaceans and influences the growth of the animal. The biogenic amines serotonin and melatonin are proved to be involved in promoting growth in crustaceans [29]. The opioid peptides leucine enkephalin and methionine enkephalin produced from the crustacean eyestalk also play a role in molt regulation. Especially leucine enkephalin involved in promoting growth in crab *Oziotelphusa senex senex* [30]. Besides this temperature of the medium and photoperiod also promote molting in some crustaceans [31-35]. However, the role of biogenic amines, opioid peptides and external factors are still under dearth of literature, need more attention of researchers [36-45].

Conclusion

It is essential to emphasis that food scarcity, in particular protein is the biggest hurdle standpoint in the World. It is essential to look for alternatives to produce more food within the available resources [46-51]. Crustacean aquaculture produces, high quality and delicious protein, but has its own limitations to produce added amount of protein. To produce crustacean protein, further alternatives are in search. This review focused on the potential lacunas in crustacean molting research. Such as

- A. Controlling of molt inhibitory gene expression by means of RNAi technique
- B. Growth increase by using appropriate Probiotic and Probiotics which can induce ecdysterodiogenesis in Y-organs and MF synthesis in MOs respectively
- C. Focused studies desirable on biogenic amines, options and external factors [52-58].

Acknowledgement

The authors are thankful to University Grants Commission, New Delhi for supporting Dr. P. Ramachandra Reddy with UGC-Raman Post-Doctoral Fellowship in USA (UGC Lr. F. NO.5-85/2016(IC)) for the year 2016-17.

References

1. Varalakshmi K, Ravichandra Reddy (2010) Effects of eyestalk ablations on growth and ovarian maturation of the freshwater prawn *Macrobrachium lanchesteri* (de Man) Turk J Fish Aquatic Sci 10: 403-410.
2. Reddy PS, Ramamurthi R (1999) Recent trends in crustacean endocrine research. PINSAs 15-32.
3. Reddy PR, Reddy PS (2006) Isolation of peptide hormones with pleiotropic activities in the freshwater crab, *Oziotelphusa senex senex*. Aquaculture 259(1-4): 424-431.

4. Reddy PR, Nagaraju GPC, Reddy PS (2004) Involvement of methyl farnesoate in the regulation of molting and reproduction in the freshwater crab *Oziotelphusa senex senex*. *J Crust Biol* 24(3): 511-515.
5. Venkitraman PR, Jayalakshmy KV, Balasubramanian T (2010) Effect of eyestalk ablation on molting and growth in penaeidprawns: *Metapenaeus monoceros*. *Indian J Fish* 57(2): 25-32.
6. Mohamed A Amer, Awaad AM, El Sayed, Khalid A, Al Damhougy, et al. (2015) Inducing Molting by Eyestalk Ablation in the Red Swamp Crayfish, *Procambarus clarkii* from the Egyptian Freshwaters. *Am J Life Sci* 3(6-1): 69-75.
7. Rotllant G, Takac P, Liu L, Scott GL, Laufer H (2000) Role of ecdysteroids and methyl farnesoate in morphogenesis and terminal moult in polymorphic males of the spider crab, *Libinia emarginata*. *Aquaculture* 190(1-2): 103-118.
8. Neelima H, Reddy PR, Reddy PS (2016) Natural and Induced (Eyestalk Ablation) Molt Cycle in Freshwater Rice Field Crab *Oziotelphusa Senex Senex*. *J Aquac Res Development* 7: 424.
9. Okumura T, Aida K (2001) Effects of bilateral eyestalk ablation on molting and ovarian development in the giant freshwater prawn, *Macrobrachium rosenbergii*. *Fish Sci* 67(6): 1125-1135.
10. Chung JS, Webster SG (2003) Molt cycle-related changes in biological activity of moult-inhibiting hormone (MIH) and crustacean hyperglycemic hormone (CHH) in the crab, *Carcinus maenas*. *Eur J Biochem* 270(15): 3280-3288.
11. Chen S, Huner JV, Malone RF (1993) Molting and mortality of red swamp and white river crawfish subjected to eyestalk ablation: A preliminary study for commercial soft-shell crawfish production. *J World Aquacult Soc* 24(1): 48-57.
12. Majid AH, Nader S, Azadeh A, Ahmad B (2008) Influence of eyestalk ablation and temperature on molting and mortality of narrow-clawed crayfish (*Astacuseptodactylus*). *Turk J Fish Aquatic Sci* 8: 219-223.
13. Keller R (1992) Crustacean neuropeptides: Structures, functions and comparative aspects. *Experientia* 48(5): 439-448.
14. Huberman A (2000) Shrimp endocrinology: A review. *Aquaculture* 191(1-3): 191-208.
15. Diwan AD (2005) Current progress in shrimp endocrinology. *Indian J Exp Biol* 43(3): 209- 223.
16. Chang ES, Chang SA, Mulder EP (2001) Hormones in the Lives of Crustaceans: An Overview, *Amer Zool* 41: 1090-1097.
17. Nagaraju GPC, Prasad GLV, Reddy PS (2005) Isolation and characterization of mandibular organ - inhibiting hormone from the eyestalks of freshwater crab, *Oziotelphusa senex senex*. *International Journal of Applied Science and Engineering* 3(1): 61-68.
18. Swetha CH, Sainath SB, Reddy PR, Reddy PS (2011) Reproductive endocrinology of female crustaceans: perspective and prospective. *J Marine Sci Res Develop* S3-S13.
19. Sarojini R, Nagabhushanam R, Fingerman M (1995) A neurotransmitter role for red-pigment concentration hormone on ovarian maturation in the red swamp crayfish *Procambarus clarkii*. *J Exp Biol* 198(6): 1253-1257.
20. Chung SJ (2010) Hemolymph ecdysteroids during the last three molt cycles of the blue crab, *Callinectes sapidus*: quantitative and qualitative analyses and regulation. *Arch Insect Biochem Physiol* 73(1): 1-13.
21. Gatesoupe FJ (2007) Live yeasts in the gut: natural occurrence, dietary introduction, and their effects on fish health and development. *Aquaculture* 267(1-4): 20-30.
22. Qi Z, Zhang XH, Boon N, Bossier P (2009) Probiotics in aquaculture of China: current state, problems and prospect. *Aquaculture* 290(1-2): 15-21.
23. Ringø E, Zhou Z, Vecino JLG, Wadsworth S, Romero J, et al. (2015) Effect of dietary components on the gut microbiota of aquatic animals. A never-ending story? *Aquacul Nutrition* 22(2): 219-282.
24. Laufer H, Landau M, Borst D, Homola E (1986) The synthesis and regulation of methyl farnesoate, a new juvenile hormone for crustacean reproduction. In: Porchet M, Andries JC, Dhainaut A (Eds.), *Advances*.
25. Wainwright G, Prescott MC, Rees HH, Webster SG (1996) Mass spectrometric determination of methyl farnesoate profiles and correlation with ovarian development in the edible crab, *Cancer pagurus*. *J Mass Spectrom* 31(12): 1338-1344.
26. Rodriguez EM, Medesani DA, Lopez Greco LS, Fingerman M (2002) Effects of some steroids and other compounds on ovarian growth of the red swamp crayfish, *Procambarus clarkii*, during early vitellogenesis. *J Exp Zool* 292(1): 82-87.
27. Suneetha Y, Naga Jyothi P, Reddy MS (2010) Impact of methyl farnesoate in the regulation of molting and reproduction in the tropical penaeid prawn, *Penaeus monodon*. *Glob J Biotech Biochem* 5(4): 226-230.
28. Hemalatha M, Parameswari K, Kishori B, Reddy BS, Reddy PR (2016) Methyl farnesoate induced ovarian maturation in freshwater prawn, *Macrobrachium rosenbergii*. *J Oceanogr Mar Res* 4(1): 1-6.
29. Sainath B, Reddy PS (2010) Evidence for the involvement of selected biogenic amines (serotonin and melatonin) in the regulation of molting of the edible crab, *Oziotelphusa senex senex Fabricius*. *Aquaculture* 302(3-4): 261-264.
30. Kulkarni GK, Glade L, Fingerman M (1991) Oogenesis and effects of neuroendocrine tissues on in vitro synthesis of protein by the ovary of the red swamp crayfish *Procambarus clarkii* (Girard). *J Crust Biol* 11(4): 513-522.
31. Pervaiz Ahmed Pervaiz, Madhu Sudan, Susan Manohar (2015) Studies on the effect of photoperiodism and temperature on moulting of a freshwater prawn *Macrobrachium dayanum*. *Inter J Fish Aquatic Studies* 3(1): 325-328.
32. Aguilar MB, Quackenbush LS, Hunt DT, Shabinowitz J, Huberman A (1992) Identification, purification and initial characterization of the vitellogenesis inhibiting hormone from the Mexican crayfish, *Procambarus bouvierii*. *Comp Biochem Physiol* 102(3): 491-498.
33. Hoang T, Matteo B, Lee SY, Clive PK, Gay EM (2003) Influence of light intensity and photoperiod on moulting and growth of *Penaeus merguensis* cultured under laboratory conditions. *Aquaculture* 216 (1-4): 343- 354.
34. Borst DW, Ogan J, Tsukimura B, Claerhout T, Holford KC (2001) Regulation of the crustacean mandibular organ. *Am Zool* 41(3): 430-441.
35. Hesni AM, Shabanipour N, Atabati A, Bitraf A (2008) Influence of eyestalk ablation and temperature on moulting and mortality of narrow-clawed Crayfish (*Astacuseptodactylus*). *Turkish Journal of Fisheries and Aquatic Science* 8: 219-223.
36. Marcela C, Montagna (2011) Effect of temperature on the survival and growth of fresh water prawns *Macrobrachium borellii* and *Palaemonetes argentinus* (Crustacea, Palaemonidae). *Iheringia Sér Zool* 101(3): 233-238.
37. Aktas M, Kumlu M, Eroldogan OT (2003) Off season maturation and spawning of *Penaeus semisulcatus* by eyestalk ablation and/or temperature-photoperiod regimes. *Aquaculture* 228(1-4): 361-370.
38. Chang ES, Mykles DL (2011) Regulation of crustacean molting: A review and our perspectives. *Gen Comp Endocrinol* 172 (3): 323-330.
39. Chang ES (1993) Comparative endocrinology of molting and reproduction. *Ann Rev Entomol* 38: 161-180.
40. Choy SC (1987) Growth and Reproduction of eyestalk ablated *Penaeus canaliculatus* (Oliver, 1811) (Crustacea: Penaeidae). *J Exp Mar*

Biol Ecol 112(2): 93-107.

41. De Kleijn DPV, Van Herp F (1998) Involvement of the hyperglycemic neuro hormone family in the control of reproduction in decapod crustaceans. *Invert Reprod Dev* 33(2-3): 263-272.
42. Jeon JM, Kim BK, Lee JH, Kim HJ, Kang CK, et al. (2012) Two type I crustacean hyperglycemic hormone (CHH) genes in *Morotoga* shrimp (*Pandalopsis japonica*): cloning and expression of eyestalk and pericardial organ isoforms produced by alternative splicing and a novel type I CHH with predicted structure shared with type II CHH peptides. *Comp Biochem Physiol B Biochem Mol Biol* 162(4): 88-99.
43. Kang BJ, Okutsu T, Tsutsui N, Shinji J, Bae SH, et al. (2014) Dynamics of vitellogenin and vitellogenesis-inhibiting hormone levels in adult and sub adult white leg shrimp, *Litopenaeus vannamei*: relation to molting and eyestalk ablation. *Biol Reprod* 90(1): 12.
44. Keller R, Sedlmeier D (1998) A metabolic hormone in crustaceans: the hyperglycemic neuro peptide. In: Lufer H, Downer RGH (Eds.), *Endocrinology of selected invertebrate types*. Volume 2, AR Liss, New York, USA, pp. 315-326.
45. Kishori B, Sreenivasula Reddy P (2003) Influence of Leucine-Enkephalin on Moulting and Vitellogenesis in the Freshwater Crab, *Oziotelphusa senex senex* (Fabricius, 1791)(Decapoda, Brachyura). *Crustaceana* 76(11): 1281-1290.
46. Lachaise F, Goudeau M, Hetru C, Kappler C, Hoffmann JA (1981) Ecdysteroids and ovarian development in the shore crab *Carcinus maenas*. *Hoppe-Seyler's Z Physiol Chem* 362(5): 521-529.
47. Lu W, Wainwright G, Webster SG, Rees HH, Turner PC (2000) Clustering of mandibular organ-inhibiting hormone and moultinhibiting hormone genes in the crab, *Cancer pagurus*, and implications for regulation of expression. *Gene* 253(2): 197-207.
48. Luo X, Chen T, Zhong M, Jiang X, Zhang L, et al. (2015) Differential regulation of hepato pancreatic vitellogenin (VTG) gene expression by two putative molt-inhibiting hormones (MIH1/2) in Pacific white shrimp (*Litopenaeus vannamei*). *Peptides* 68: 58-63.
49. Nagaraju GPC, Reddy PR, Reddy PS (2006) In vitro methyl farnesoate secretion by mandibular organs isolated from different molt and reproductive stages of the crab *Oziotelphusa senex senex*. *Fish Sci* 72(2): 410-414.
50. Nagaraju GPC (2011) Reproductive regulators in decapod crustaceans: an overview *J Exp Biol* 214 (Pt 1): 3-16.
51. Nakatsuji T, Chi-Ying Lee, Watson RD (2009) Crustacean molt-inhibiting hormone: Structure, function, and cellular mode of action. *Comp Biochem Physiol A* 152(2): 139-148.
52. Quackenbush LS (2001) Yolk synthesis in the marine shrimp, *Penaeus vannamei*. *Amer Zool* 41(3): 458-464.
53. Santos EA, Nery LEM, Keller R, Goncalves AA (1997) Evidence for the involvement of the crustacean hyperglycemic hormone in the regulation of liquid metabolism. *Physiol Zool* 70(4): 415-420.
54. Tamone SL, Prestwich GD, Chang ES (1997) Identification and characterization of methyl farnesoate binding proteins from the crab, *Cancer magister*. *Gen Comp Endocrinol* 105(2): 168-175.
55. Techa S, Chung JS (2015) Ecdysteroids Regulate the Levels of Molt-Inhibiting Hormone (MIH) Expression in the Blue Crab, *Callinectes sapidus*. *PLoS One* 10 (4): 1-19.
56. Watson RD, Spaziani E, Bollenbacher WE (1989) Regulation of ecdysone biosynthesis in insects and crustaceans: a comparison; In *Ecdysone: J Koolman Stuttgart, Georg Thieme Verlag* (Ed.), *From Chemistry to Mode of Action*, pp. 188-203.
57. Webster SG, Keller R, Dircksen H (2012) The CHH-superfamily of multifunctional peptide hormones controlling crustacean metabolism, osmoregulation, moulting, and reproduction. *Gen Comp Endocrinol* 175(2): 217-233.
58. Zmora N, Trant J, Zohar Y, Chung JS (2009) Molt-inhibiting hormone stimulates vitellogenesis at advanced ovarian developmental stages in the female blue crab, *Callinectes sapidus* 1: an ovarian stage dependent involvement. *Saline Systems* 5: 7.



This work is licensed under Creative Commons Attribution 4.0 License
DOI: [10.19080/JETR.2017.01.555556](https://doi.org/10.19080/JETR.2017.01.555556)

**Your next submission with Juniper Publishers
will reach you the below assets**

- Quality Editorial service
- Swift Peer Review
- Reprints availability
- E-prints Service
- Manuscript Podcast for convenient understanding
- Global attainment for your research
- Manuscript accessibility in different formats
(Pdf, E-pub, Full Text, Audio)
- Unceasing customer service

Track the below URL for one-step submission

<https://juniperpublishers.com/online-submission.php>