

Lipid Peroxidation, Enzymatic and Non-Enzymatic Alterations of DCM-F of *Rhizophora mucronata* in Diabetic Rats



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Abstract

Oxidative stress is responsible for impairment of β -cells caused by chronic glucose toxicity. The present study demonstrates dichloromethane fraction (DCM-F) of *Rhizophora mucronata* mediated fortification against diabetes mellitus induced alterations in antioxidant defense system in the animal model. Single intraperitoneal injection of streptozotocin and nicotinamide was to induce diabetes in rats. 50mg/kg of DCM-F was orally treated to diabetic rats for 45 days. At the end of the experiment, blood glucose, lipid hydro-peroxide (LH), plasma enzymatic and non-enzymatic antioxidants were determined. Treatment of DCM-F to the experimental rats notably ($p < 0.01$) restored blood-glucose, body weight, lipid profile and carbohydrate metabolic enzyme activities. Besides, the intensity of LH increased and CAT, glutathione peroxidase (GPx), GSH, and SOD were considerably decreased in diabetic rats. These unfavourable alterations were inverted to normal in DCM-F treated rats. Moreover, notable results on ceruloplasmin, ascorbic acid, and tocopherol were observed in DCM-F treatment as differed with diabetic and control. In conclusion, DCM-F of *R. mucronata* act as antioxidant linked with anti-hyperglycemic and act as a ligand for selected antioxidant receptors.

Keywords: Antioxidant; DCM-F; Glutathione peroxidase; Oxidative stress, *Rhizophora mucronata*

Abbreviations: CAT: Catalase; DCM-F: Dichloromethane Fraction; GPx: Glutathione Peroxidase; GSH: Glutathione; LH: Lipid Hydroperoxide; MDA: Malondialdehyde; PUFA: Polyunsaturated Fatty Acids; SOD: Superoxide Dismutase; TBARS: Thiobarbituric Acid Reactive Substances

Introduction

Free radicals produced during regular metabolism are removed by way of an efficient scavenging system and the imbalance effects in expanded oxidative strain. Lipid peroxide stages in diabetes are extended in plasma, serum, kidney, lens and in erythrocyte membrane [1]. Significant modifications in lipid metabolism and structural modifications in cell membranes are related to the progress of metabolic disorders [2]. The dysfunction among enzymatic and non-enzymatic oxidation of lipids in vivo is not always absolute [3]. Oxidative pressure has these days been proven accountable, as a minimum in the component, for pancreatic β -mobile dysfunction due to glucose toxicity. Under hyperglycemia, production of various decreasing sugars, which includes glucose-6-phosphate and fructose, will increase through glycolysis and polyol pathways [4]. During this method, reactive oxygen species (ROS) are produced and cause tissue harm. So, STZ is broadly hired to set off experimental diabetes in animals [5]. DCM-F of *Rhizophora mucronata* is confirmed anti-hyperglycemic and anti-hyperlipidemic impact on diabetic animals [6]. In the continuation of preceding research,

we have appraised the impact of DCM-F on lipid peroxidation and plasma antioxidants in STZ-NAD induced diabetic animals.

Summary

Body weight, blood glucose, cholesterol, lipid profile, plasma insulin and carbohydrate metabolic enzyme activities of experimental rats were estimated and reported in the previous studies [6]. Figure 1 illustrated the level of LH and TBARS in the plasma of experimental rats. TBARS concentrations were particularly raised in the diabetic rats, distinguished with control and DCM-F treated rats. DCM-F (50mg/kg) showed a convincing decrease in TBARS level. No consistent changes were found among control and treated rats. Activities of enzymatic antioxidants were clearly giveaway in diabetic rats compared with control and treated rats. Lipid peroxidation is a process enhancing the oxidation of polyunsaturated fatty acids, which causes metabolic disorders, cancer, and cardiovascular diseases. The results of the present study show STZ induced free radicals and increased oxidation of PUFA in the plasma and tissues of diabetic rats. This confirms the previous reports on the

proficiency of STZ to assist production of free radicals and causes oxidative stress [7]. The increase of reactive oxygen species (ROS) from the reaction of enzymes, metabolism of xenobiotics shows the way to lipid peroxidation with consecutive cell disruption and toxicity [8]. Free radical increases in liver and kidney tissues may accommodate almost absorption of per-oxidize-

able fatty acids. DCM-F treatments reduce MDA, an indicator of lipid peroxidation in the tissues of diabetic rats suggesting that the DCM-F have massive anti-oxidative properties. Alkaloids of mangroves have an important role as edible antioxidants for the preclusion of oxidative damages in humans [9].

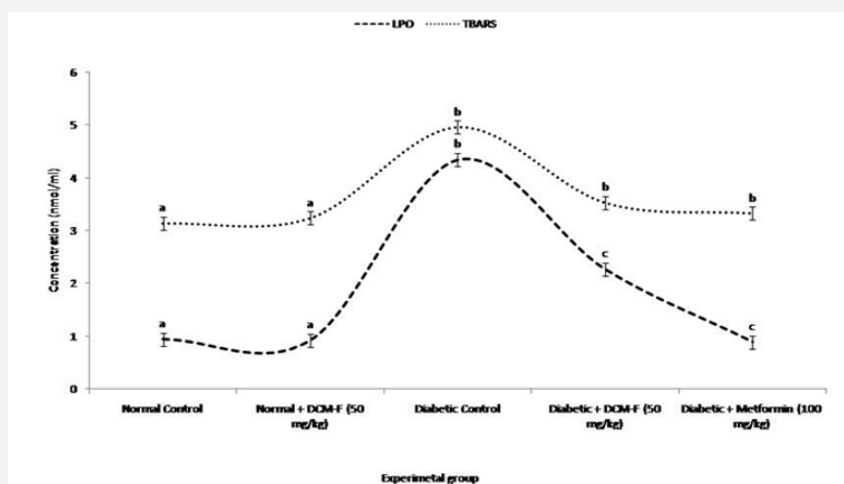


Figure 1: Effect of DCM-F on lipid hydroperoxides and TBARS levels in plasma of normal and diabetic rats. Each value is mean S.D. for 8 rats in each group (n=8). Values that have a different superscript letter (a,b,c) differ significantly with each other (P<0.05, Duncan’s multiple range test).

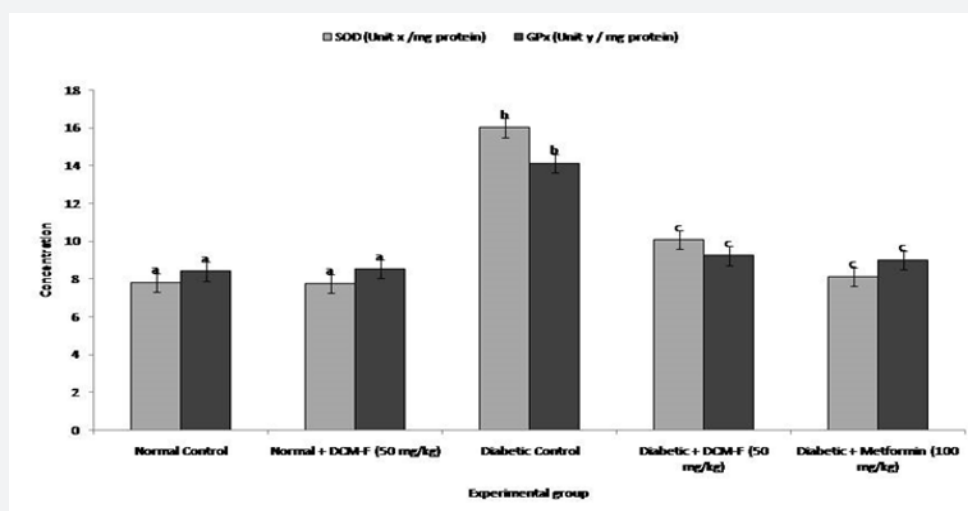


Figure 2 : Effect of DCM-F on SOD and GPx levels in plasma of normal and diabetic rats. Each value is mean S.D. for 8 rats in each group (n=8). Values that have a different superscript letter (a,b,c) differ significantly with each other (P<0.05, Duncan’s multiple range test) x-One unit of SOD is defined as the enzyme concentration which gives 50% inhibition of NBT reduction in one minute; y- One unit of GPx is defined as the μ g of glutathione consumed per minute.

DCM-F treatment explained a rational (P<0.05) get higher in the actions of superoxide dismutase, catalase (Table 1), and glutathione peroxidase (Figure 2). Notable (P<0.05) increase in the levels of glutathione, ascorbic acid, α-tocopherol and ceruloplasmin in diabetic rats contrasted to control. DCM-F treatment progress to a significant (P<0.05) increase in the plasma concentrations of glutathione, ascorbic acid, α-tocopherol, and ceruloplasmin if compared with diabetic

control rats (Table 1). No consistent variations were found among control and treated rats. Catalase and superoxide dismutase plays a major role in the reduction of highly reactive hydroxyl radicals and dismutation of superoxide radicals respectively [10-12]. Glutathione peroxidase detoxifies hydrogen-peroxide into water through the oxidation of reduced glutathione [13,14]. The present study results confirm the restored action of enzymatic antioxidants through the treatment of DCM-F. In the

continuation of lipid peroxidation and formation of hydrogen peroxides are arisen. Inhibition of hydrogen peroxide means that production of hydroxyl radicals is reduced which protects the cells from xenobiotics [15]. In this view, antioxidant alkaloids reduced the oxidative damages through inhibition of free radical formation [16]. Depletion of glutathione may be accompanying to the increased lipid peroxidation in the STZ induced diabetic rats. Previous reports express decreased antioxidant enzyme action enhanced peroxidative status particularly liver and kidney tissues of diabetic rats [17,18]. During peroxidation, α -tocopherol reduces lipid hydroperoxides and protects cell

damage, while binding to the copper ion ceruloplasmin inhibits lipid peroxidation [19]. Increased utilization as an antioxidant defense system and reduced in the level of glutathione in diabetic rats reduced ascorbic acid [20]. These are supports, DCM-F of *Rhizophora mucronata* treatment restored antioxidant enzyme action in diabetic rats through detoxification and scavenging of free radicals. *Rhizophora mucronata*, the mangrove plant isolated DCM-F was studied to control diabetes mellitus to enhance the action of antioxidant enzymes. In conclusion, DCM-F of *Rhizophora mucronata* act as antioxidant linked with anti-hyperglycemic effect.

Table 1: Effect of DCM-F on CAT, GSH, Ceroplasmin, Ascorbic acid, and α -tocopherol levels in plasma of normal and diabetic rats.

Experiments	Normal Control	Normal Rat + DCM-F (50 mg/kg)	Diabetic Control	Diabetic Rat + DCM-F (50 mg/kg)	Diabetic Rat + Metformin (100 mg/kg)
CAT (unit y/ mg protein)	0.52 ± 0.22 ^a	0.57 ± 0.31 ^a	3.79 ± 0.57 ^b	0.68 ± 0.61 ^c	0.94 ± 0.18 ^c
GSH (mg/dL)	93.72 ± 2.01 ^a	90.15 ± 2.11 ^a	28.51 ± 1.79 ^b	91.11 ± 2.10 ^c	85.2 ± 2.14 ^c
Ceroplasmin (nmol/l)	1.39 ± 0.10 ^a	1.40 ± 0.11 ^a	0.85 ± 0.06 ^b	1.26 ± 0.07 ^c	1.32 ± 0.31 ^c
Ascorbic acid (nmol/l)	0.13 ± 0.02 ^a	0.12 ± 0.01 ^a	0.05 ± 0.02 ^b	0.10 ± 0.02 ^c	0.11 ± 0.01 ^c
α -tocopherol (nmol/l)	0.05 ± 0.003 ^a	0.05 ± 0.002 ^a	0.02 ± 0.001 ^b	0.04 ± 0.002 ^c	0.05 ± 0.001 ^c

Each value is mean S.D. for 8 rats in each group (n=8).

Values that have a different superscript letter (a,b,c) differ significantly with each other (P<0.05, Duncan's multiple range test).

z-One unit of CAT is defined as the μ mole of hydrogen peroxide consumed per minute.

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