The Gut: A Key to the Pathogenesis of Type 2-Diabetes?

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Mini Review

The gastrointestinal tract plays a predominant role in the regulation of the postprandial plasma glucose levels [1]. The first regulating factor is the gastric antral motility which, by incompletely elucidated neuroendocrine mechanisms, regulates the rate of emptying and hence exposure of chyme to the small intestine. Thus, it is a main function of the stomach to receive and retain the incoming meal, and eventually to expel the triturated, emulsified and partly digested contents, the chyme, at a slow and surprisingly constant rate of maximally 4Kcal per minute. The duration of gastric emptying thereby becomes regulated by the energy content of the ingested meal [2]. The exposure rate of the small intestine is similarly limited and relatively constant. If the regulatory mechanisms are interfered with (for instance with surgical procedures like pyloroplasty or gastric bypass) and the normal emptying rate is exceeded, a so-called “dumping syndrome” may be elicited.

This syndrome which includes nausea, malaise, desire to lie down, maybe fainting, is caused by the increased osmotic load presented to the small intestine which shifts fluid from the circulation to the intestinal lumen via the leaky epithelium in the proximal gut. The more or less constant emptying rate also means that the secretion and thereby the plasma concentration of some of the gut hormones, e.g. the insulin-stimulating hormone, glucose-dependent insulin tropic polypeptide (GIP) which is dependent on the intestinal absorption rate of nutrients, quickly rises to a certain elevated level which is then maintained as long as there is emptying from the stomach, depending on the total amount of stimulatory nutrients that were present in the meal (and are being retained in the stomach) [2].

It is possible that hydrogen ions in the duodenum (released from the stomach) play a role in the regulation of the emptying rate, but nutrients in the duodenum may also inhibit the emptying rate by neuroendocrine mechanisms. The hormone, GIP, has little effect on gastric emptying, whereas the other incretin hormone, glucagon-like peptide-1 (GLP-1), the secretion of which follows a similar pattern, powerfully inhibits gastric emptying. There is no doubt that the regulation of gastric emptying plays a major role for the postprandial glucose responses, again as illustrated in conditions of accelerated emptying rates, where particularly the early postprandial glucose excursions may be dramatically increased [3]. The increased secretion of GIP and GLP-1 powerfully amplifies the glucose-induced insulin secretion, and thereby also the ability of the organism to limit and restore to normal the increased glucose concentrations—indeed postprandial reactive hypoglycemia (sometimes designated “late dumping”) may ensue, and is relatively often observed in individuals with accelerated gastric emptying [4].

The amplifying effect of the gut hormones on the glucose-induced insulin secretion is designated the incretin effect, and this is responsible for the proportionate increase in insulin secretion according to the ingested amount of glucose (and other meal components). The effects is small after small meals (small amount of glucose), but increases so that it may be responsible for up to 80% of the elimination of glucose from the circulation after larger meals (corresponding to 100g of glucose) [2,5]. By means of the regulation of gastric emptying and the incretin effect, it is ensured that postprandial glucose excursions are normally moderate and in fact independent of the ingested amount of calories (glucose) [1].

It is likely that these mechanisms are of major importance to prevent complications elicited by hyperglycemia. As mentioned these powerful mechanisms may also lead to severe postprandial reactive hypoglycemia [6]. The question then arises whether the same mechanisms play a role in the development of type 2 diabetes mellitus (T2DM)? The gastric emptying rates in patients with T2DM show great variability and a consistent
abnormality do not seem to prevail (although abnormalities may very well exist in selected groups).

The early response (0-60min) to ingestion of a mixed meal of the two hormones, GIP and GLP-1, does not seem to be importantly affected in patients with T2D, but GLP-1 secretion is more consistently impaired in the later phase (from 60 minutes and onwards) [7] and this may play a significant role. A similar impairment (and a similar impairment of the incretin effect, see below) is seen in obesity [8,9]. Probably it is of greater importance that the insulin tropic action of both hormones is also impaired. During the progressive development of glucose intolerance, a gradual loss of the incretin effect, as well as the insulin tropic action of the two hormones are observed [10] and their effect is almost completely lost in full-blown diabetes as far as physiological amounts of the hormones are concerned [11].

However, in contrast to the almost complete loss of insulin tropic action of GIP, regardless of dose, supraphysiological concentrations of GLP-1 may still have considerable effect [12]. Evidently, the loss of the incret effect is of major importance for the postprandial hyperglycaemia of the patients with T2D and restoration of the incretin effect with GLP-1-receptor agonists provides part of the explanation for the antidiabetic effects of these agents. The GLP-1-receptor agonists also powerfully inhibits the gastric emptying rate, at least acutely and this explains part of the effect on postprandial glycaemia of the short-acting GLP-1-receptor agonists (eventide and lixisenatide).

Regarding the long-acting GLP-1-receptoragonists, tachyphylaxis rapidly develops regarding their effects on gastric emptying, while their incretin effect is preserved and when during meal intake the plasma glucose concentration rises, they will exert their incretin action, which at the cellular level in essence consists of potentiating of the glucose-induced insulin secretion [13,14]. There is, however, an additional factor which plays a rather important role: patients with T2DM typically have increased plasma concentrations of glucagon, both in the fasting state and postprandial [7].

There is a general misconception that meal intake should lead to inhibition of glucagon secretion; on the contrary, most meals, in particular protein-rich meals, will stimulate glucagon secretion, but for patients with T2DM the increase is even larger. Because glucagon stimulates the hepatic glucose production, this rise has considerable consequences for the postprandial plasma profiles) superimposed on an intravenous glucose infusion mimicking the concentration curves resulting from an OGTT [16]. Individually, it turned out, each of the hormones had the mentioned, expected effects, but infused together the resulting glucagon secretion profile was similar to that observed after oral glucose alone. Particularly GIP stimulated glucagon secretion. An increased activity of GIP might therefore contribute to the postprandial hyperglucagonemia in T2DM. This together with the loss of its insulin tropic effect in T2DM might therefore suggest that GIP actually has diabetogenic effects [17].

However, it is also possible that the postprandial hyperglucagonemia is derived not from the pancreas but from the GI-tract where some of the endocrine cells, under certain circumstances, which may include T2DM, may be able to produce glucagon. This is clearly seen in individuals after total pancreatectomies, who exhibit a large postprandial glucagon response after oral glucose [18]. It can actually be demonstrated that the response has important effects for hepatic glucose production.

Extra pancreatic glucagon may therefore contribute to the diabetic hyperglucagonemia. Furthermore, there is strong evidence from recent research that circulating amino acids play a prominent role in the regulation of glucagon secretion, and in view of this, it should always be suspected that altered amino acid levels underlie clinical hyperglucagonemia. The reverse also seems to be true: that the plasma level of glucagon plays a predominant role in the regulation of the plasma levels of the amino acids; thus conditions with hyperammonaemia are associated with low and high levels of plasma amino acids, respectively [19]. Indeed, amino acid metabolism and the glucagon-producing alpha cells in the pancreas seem to be coupled in a close, negative feedback loop. Most recently we examined patients with varying degrees of non-alcoholic fatty lived disease and found a correlation between on one hand liver function and amino acid levels and on the other hand the plasma glucagon levels [20].

The hypothesis is that the steatosis impairs liver function which also comprises an impairment of the effects of glucagon on amino acid metabolism and urea genesis. As a consequence, plasma levels of amino acids rise, which in turn results in hyperglucagonemia. Because glucagon also influences glucose metabolism and hepatic glucose production, this will also result in glucose intolerance and increased postprandial glucose excursions. A lot of work still has to be done with respect to unravelling these associations but the recent application of glucagon receptor antagonists and the use of transgenic animals have greatly facilitated these studies in both people and experimental animals Figure 1 [21].
One final factor deserves discussion: there is actually a fasting level of the hormone GLP-1, and it can be demonstrated, also in humans, that blocking of the GLP-1 receptor with a GLP-1 receptor antagonist (extending 9-39) results in increased glucagon secretion, decreased insulin secretion and presumably as a consequence of this, glucose intolerance and hyperglycemia in the fasting state [22]. As already mentioned, the GLP-1 secretion is often impaired in T2DM, perhaps mainly related to simultaneously occurring obesity [8]. Because the pathogenesis of T2DM also is associated with the development of obesity, a picture emerges where decreased GLP-1 secretion leads to both an impaired incretin effect and increased glucagon secretion, both with diabetogenic actions. As hepatic steatosis develops because of increased calorie intake, the liver’s sensitivity to both insulin and to the actions of glucagon on amino acid and glucose metabolism will be impaired. Furthermore, parallel development of insulin resistance (because of ectopic fat) in the skeletal muscles results in impaired peripheral glucose uptake. Together, all of these result in a vicious diabetogenic cycle.

References


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