



Combination Of Oncolytic Newcastle Disease Virus (Ndv) and Vaccine Vector Adenovirus (Adv) as a Potential Virotherapy for Cancer: A Systematic Review



Ferbian Milas Siswanto^{1*} and Pakajiraporn Arunngam²

¹Department of Biotechnology, Dhyana Pura University, Indonesia

²Department of Biomedical Chemistry, Kwansai Gakuin University, Japan

Submission: April 07, 2018; Published: April 13, 2018

*Corresponding author: Ferbian Milas Siswanto, Department of Biotechnology, School of Health, Science and Technology, Dhyana Pura University, Badung, Indonesia, Tel: 6281337321736; Email: ferbianms@gmail.com

Abstract

Cancer is a disease with high morbidity and mortality, one of the leading causes of death in the world. Nowadays, the foremost clinical cancer therapy in a patient are surgery, chemotherapy and/or radiotherapy. Despite of the great amount of research on cancer and advance technology in medicine, the mortality rate of cancer remain high due to limited therapeutic effects and additional side effects of current therapy. Here we provide an overview on the virotherapy using the combination of Newcastle disease virus (NDV) and the adenovirus (AdV). Both NDV and AdV possess an oncolytic activity and a potential as vector vaccine. However, oncolytic activity of NDV is more potent than adenovirus. In contrast, the AdV potential as a vector of cancer vaccines is better than NDV. Therefore, in this paper, we discuss the development of a virotherapy combination by utilizing oncolytic activity of NDV, and vaccine vector AdV simultaneously for cancer therapy to improve the effectiveness of therapy against cancer. Conclusion: Decreased estrogen level following ovariectomy causes osteoporosis.

Keywords: Newcastle disease virus; Adenovirus; Virotherapy; Cancer

Abbreviations: NDV: Newcastle Disease Virus; AdV: Adeno Virus; VVND: Velogenic Viscerotropic Newcastle Disease; PBMC: Peripheral Mononuclear Cells; HN: Hemagglutinin-Neuraminidase; TRAIL: TNF-Related Apoptotic-Inducing Ligand; JNK: c-Jun N-terminal Kinase; NOS: Nitric Oxide Synthase; dsDNA: Double-Stranded DNA; NK: Natural Killer; CAE: Carcioembryonic Antigen; TLRs: Toll like receptors

Introduction

Cancer is a disease with high morbidity and mortality that leads to death. Until now, cancer is still the leading cause of death in humans. In 2012, approximately 14.1 million cases of cancer have been reported worldwide, and have caused the deaths of 8.2 million people (about 15% of all deaths). It is characterized by uncontrolled cell division, invade surrounding tissue, and metastasize to other organs in the body. The four most commonly reported cancers are lung, breast, colon, and prostate cancer. However, all organs in the human body can be cancer regardless of age, gender, ethnicity, diet, and environment [1]. Generally, cancer is caused by decreased cell death or increased cell proliferation. In other words, any dysregulation of cell cycle or apoptosis will result in uncontrolled cell growth or malignancy [2].

Cancer occurs due to genetic and environmental factors that cause deviations in the growth regulation of stem cell populations. Improving knowledge of the molecular

processes underlying cancer development, as well as advances in diagnostic techniques, radiotherapy technology, and chemotherapy, has increased the survival rate of cancer patients. However, recent therapy has not greatly improved the survival of cancer patients who have undergone metastasis. Although modern technology has been developed, cancer is still afflicted millions of people worldwide [3]. This is because, in addition to the limited therapeutic effects, radio and chemotherapy also cause side effects [1]. The ideal cancer therapy is a therapy that selectively kills malignant cells, and does not damage other normal cells in the body. Currently, radiotherapy, chemotherapy, and surgery are the most common modalities in cancer therapy. However, these therapies often cause harmful side effects [4] and often lead to resistance [5].

Therefore, the development of cancer therapy with high effectivity and selectivity for cancer cells with minimum side effects becomes crucial. The idea of using bacteria and viruses

to treat malignancy in humans began in the mid-1800s in which tumor regression was associated with bacterial and viral infections [6]. The development of cell culture technique and virus technology in the early 1950s led researchers to learn more about the potential of viral therapy in human and small animal tumors [7]. The virus is then proven to be useful as an oncolytic agent and immunostimulator. Newcastle disease virus (NDV) that naturally infected poultry, and adenovirus (AdV) that causes human flu, is a potential viral combination as a virotherapy and immunotherapy agent. NDV can directly kill cancer cells (oncolytic activity) and adenovirus can help to stimulate the immune system to recognize cancer cells (immunostimulator activity).

Newcastle Disease Virus (NdV) as an Oncolytic Agent

Newcastle Disease Virus (NDV) is a virus of the order Mononegavirales because it has single strand RNA, negative polarization, unbranched and linear genome [8]. Furthermore, this virus occupies the family of paramyxoviridae due to its pleomorphic envelope, round-shaped with a diameter of 100-500nm, but some are in the form of filaments [9]. This virus causes Newcastle disease that attacks various poultry, especially chickens. Until now, Newcastle disease has been found in various parts of the world including Indonesia, and the cases of velogenic viscerotropic Newcastle disease (VVND) have been reported in Indonesia [10]. In Indonesia, Newcastle disease is endemic as indicated by the finding of this case throughout the year [9].

NDV was firstly reported to possess an oncolytic activity in the mid-1950s [11]. The clinical evaluation of this virus as an anticancer agent over the last few decades shows its safety and effectivity. The effectivity of NDV application is based on high oncolytic activity, and safety of its use is based on replication that selectively attacks tumor cells and does not damage normal cells. Scientists are interested in the use of NDV because it replicates more rapidly in tumor cells than normal cells in humans, and cause oncolytic effects [12]. NDV replicates 10,000 times faster in cells undergoing neoplastic changes than normal human cells in general [13,14]. There are several molecular pathways that cause the oncolytic effects of NDV, such as apoptosis pathway [1,15]. Induction of apoptosis by NDV includes a series of virus entry processes, replication, de novo protein synthesis and activation of caspases [16]. NDV induces apoptosis through both extrinsic and intrinsic pathways.

NDV-induced apoptosis is generally mediated by intrinsic pathway during the late stage of infection, while in the early stage of infection is more likely to be mediated by extrinsic pathway [17]. Activation of intrinsic pathway involves the increased activity of p53 and Bax proteins, as well as decreased expression of the Bcl-2 gene [18] which will activate the Caspase 9. The matrix protein (M protein) of NDV binds to Bax protein and increases apoptosis [19]. Whereas, the extrinsic pathway of apoptosis is induced by NDV-mediated activation of pro-apoptotic cytokines such as IFN- α and TNF- α in peripheral mononuclear

cells (PBMC) via its Hemagglutinin-Neuraminidase (HN) proteins [20,21]. The HN protein of NDV also induces expression of TNF-related apoptotic-inducing ligand receptor (TRAIL) [22,23] which further activate caspase 8 [17]. A study has shown that NDV initiates the synthesis of nitric oxide synthase (iNOS), thus increasing apoptosis via the NF κ B pathway [24,25].

NDV-infected mouse PC12 pheochromocytoma cell was proved to induce the activation of reticulum endoplasmic eIF2 α kinase (PERK) resulting in phosphorylation of eIF2 α and caspase 12 activations. Endoplasmic reticulum stress may be responsible for the activation of apoptotic pathways in cancer cells infected with NDV [26]. In addition, the induction of the external pathway by NDV also the activation of c-Jun N-terminal kinase (JNK) and p38 pathways, and decreased Akt pathway activity [27]. NDV has an excellent potential as a highly selective virotherapy candidate. This selective effect arises because of the restriction of V protein by host and secretion of virus-induced cytokines (IFN- γ and TNF- α) [28]. The first step of infection by NDV occurs in all types of cells in the body, while the second step (associated with viral replication) occurs only in tumor cells because this stage is terminated very quickly in normal cells [5]. In general, the specificity of NDV to cancer cells occurs because of damage to antiviral pathways and apoptosis in cancer cells [29].

In addition to direct cytopathic effects, NDV anti-cancer activity is associated with the activation of both innate and adaptive immune responses. NDV infection initiates the macrophage-induced synthesis of enzymes that increase anti-tumor activity in both in vitro and in vivo studies [30]. NDV stimulates monocytes that play a role in killing tumor cells via TRAIL induction [31]. The activation of natural killer (NK) cells is also involved in the cytotoxicity mediated by NDV [20]. However, to induce host immune system, the use of cancer vaccines is believed to have far more effective effects than the immunostimulator effects of NDV. The immunotherapeutic approach aims to promote the host antitumor immune response that can destroy tumor cells in both primary and metastatic-affected sites [32]. Genetic therapy-based cancer vaccination technology has been widely developed, with the virus being the most popular vector studied. Adenovirus, in addition to having oncolytic activity, is a very potential and widely used vector on cancer gene therapy and as a vaccine to express foreign antigens [33].

Adenovirus (AdV) as a Vaccine Vector

Adenovirus is a group of viruses from the Adenoviridae family responsible for 5-10% of upper respiratory infections, gastroenteritis, conjunctivitis, and cystitis (CDC, 2015). It has no envelope, icosahedral capsid with a diameter of 70-90 nm and the double-stranded DNA (dsDNA) [34]. Adenovirus has long been used as a vector for gene therapy due to its ability to influence cell biological activity, tolerate large genetic modifications, and encode proteins without integrating into the host cell genome. More specifically, the virus is used as a vector for administration

of therapeutic targets, either in the form of recombinant DNA or proteins [35].

Several studies using various antigens proved that adenovirus (AdV) is potential as a vector of cancer antigens such as glycoprotein 33 (GP33) from lymphocytic viral choriomeningitis [36], carcinoembryonic antigen (CAE) [37], beta-galactosidase antigen [38], GM-CSF antigen (such as T-VEC and Pexa-Vec) [39], E7 antigen from human papillomavirus [40], the gp100 antigen and TRP-2 antigen [41]. It may enhance cellular immunity mediated by T-cell CD8+ cells and IFN- γ -mediated humoral immune specific to cancer cells. The use of AdV as a vaccine vector is relatively safe to use with intradermal methods [42]. Adenovirus administration may stimulate ligand expression of Toll-like receptors (TLRs) and may alter cancer immunosuppressive and proinvasive microenvironment becoming proinflammatory, thus facilitating immunocompetent cells to fight against cancer [39,43,44].

General Perspective

Both NDV and AdV have oncolytic activity and potential as vector vaccine for cancer. However, oncolytic activity of NDV is more potent than adenovirus. In contrast, the AdV potential as a vector of cancer vaccines is better than NDV. Therefore, the development of a virotherapy combination by utilizing oncolytic activity of NDV, and vaccine vector AdV for cancer simultaneously are expected to improve the effectiveness of therapy against cancer. The use of an appropriate combination ratio of these two agents will improve their therapeutic potential for cancer [45,46].

References

- Ravindra PV, Tiwari AK, Ratta B, Bais MV, Chaturvedi U, et al. (2009) Time course of Newcastle disease virus-induced apoptotic pathways. *Virus Res* 144(1-2): 350-354.
- Lowe SW, Lin AW (2000) Apoptosis in cancer. *Carcinogenesis* 21: 485-495.
- Al-Qubaisi M, Rozita R, Yeap SK, Omar AR, Ali AM, et al. (2011) Selective cytotoxicity of goniiothalamine against hepatoblastoma HepG2 cells. *Molecules* 16(4): 2944-2959.
- Blomqvist C, Elomaa I, Rissanen P, Hietanen P, Nevasaari K, et al. (1996) FEC (5-fluorouracil-epirubicin- cyclophosphamide) monthly versus FEC weekly in metastasis breast cancer. First results of a randomized trial. *Acta Oncol* 31(2): 231-236.
- Fournier P, Achirmmacher V (2013) Oncolytic Newcastle Disease Virus as Cutting Edge between Tumor and Host. *Biology* 2(3): 936-975.
- Dock G (1904) The influence of complicating diseases upon leukemia. *Am J Med Sci* 127(4): 563-592.
- Kelly E, Russell SJ (2007) History of oncolytic viruses: genesis to genetic engineering. *Mol Ther* 15(4): 651-659.
- Alexander DJ (2000) Newcastle disease and other avian paramyxoviruses. *Rev Sci Tech Off Int Epiz* 19(2): 443-462.
- Kencana GAY, Kardena IM, dan Mahardika IG NK (2012) Peneguhan Diagnosis Penyakit Newcastle Disease Lapang pada Ayam Buras di Bali Menggunakan Teknik RT-PCR. *Jurnal Kedokteran Hewan* 6(1): 28-31.
- Adi AAAM, Kardena IM, Astawa NM, Putra KSA, Hayashi Y (2011) Kloning, Sikuensing dan Analisis Filogenetik Gen Nukleokapsid Protein Virus Tetelo Isolat Bali-1/07. *J Vet* 12(3): 173-179.
- Flanagan AD, Love R, Tesar W (1955) Propagation of Newcastle disease virus in Ehrlich ascites cells in vitro and in vivo. *Proc Soc Exp Biol Med* 90(1): 82-86.
- Nelson NJ (1999) Scientific interest in Newcastle disease virus is reviving. *J Natl Cancer Inst* 91(20): 1708-1710.
- Schirmmacher V, Haas C, Bonifer R, Ahlert T, Gerhards R, et al. (1999) Human tumor cell modification by virus infection: an efficient and safe way to produce cancer vaccine with pleiotropic immune stimulatory properties when using Newcastle disease virus. *Gene Therapy* 6(1): 63-73.
- Pecora AL, Rizvi N, Cohen GI, Meropol NJ, Serman D, et al. (2002) Phase I trial of intravenous administration of PV701, an oncolytic virus, in patients with advanced solid cancers. *J Clin Oncol* 20(9): 2251-2266.
- Molouki A, Hsu YT, Jahanshahi F, Rosli R, Yusoff K (2010) Newcastle disease virus infection promotes Bax redistribution to mitochondria and cell death in HeLa cells. *Intervirology* 53(2): 87-94.
- Ravindra PV, Tiwari AK, Ratta B, Chaturvedi U, Palia SK, et al. (2008) Induction of apoptosis in Vero cells by Newcastle disease virus requires viral replication, de-novo protein synthesis and caspase activation. *Virus Res* 133(2): 285-290.
- Elankumaran S, Rockemann D, Samal SK (2006) Newcastle disease virus exerts oncolysis by both intrinsic and extrinsic caspase-dependent pathways of cell death. *J Virol* 80(15): 7522-7534.
- Ravindra PV (2008) Elucidation of apoptotic pathways induced by Newcastle disease virus in cultured cells and assessment of its oncolytic potential in experimentally induced tumor. *Indian Veterinary Research Institute (Deemed University), Izatnagar, Uttar Pradesh, India*, pp. 1-135.
- Molouki A, Yusoff K (2012) NDV-induced apoptosis in absence of Bax; evidence of involvement of apoptotic proteins upstream of mitochondria. *Virol J* 9: 179.
- Zorn U, Dallmann I, Grosse J, Kirchner H, Poliwoda H, et al. (1994) Induction of cytokines and cytotoxicity against tumor cells by Newcastle disease virus. *Cancer Biother* 9(3): 225-235.
- Mi ZQ, Jin NY, Sun YC, Li X, Lian H, et al. (2004) Anti-tumour research on mouse melanoma with combined application of Newcastle disease virus and its HN gene. *Ai Zheng* 23: 910-913.
- Batliwalla FM, Bateman BA, Serrano D, Murray D, Macphail S, et al. (1998) 15-year follow-up of AJCC stage III malignant melanoma patients treated postsurgically with Newcastle disease virus (NDV) oncolysate and determination of alterations in the CD8 T cell repertoire. *Mol Med* 4(12): 783-794.
- Kumar R, Tiwari AK, Chaturvedi U, Kumar GR, Sahoo AP, et al. (2012) Velogenic newcastle disease virus as an oncolytic virotherapeutics: in vitro characterization. *Appl Biochem Biotechnol* 167(7): 2005-2022.
- Ten RM, Blank U, LeBail O, Kourilsky P, Israel A (1993) Two factors IRF-1 and KBF1/NFkB cooperate during induction of MHC class I gene expression by interferon alpha beta or Newcastle disease virus. *Compt Rend Acad Sci* 316(5): 496-501.
- Hrabak A, Csuka I, Bajor T, Csatory LK (2006) The cytotoxic effect of MTH-68/H, a live attenuated Newcastle disease virus is mediated by the induction of nitric oxide synthesis in rat peritoneal macrophages in vitro. *Cancer Lett* 231(2): 279-89.
- Fabian Z, Csatory CM, Szeberenyi J, Csatory LK (2007) p53-independent endoplasmic reticulum stress-mediated cytotoxicity of a Newcastle disease virus strain in tumor cell lines. *J Virol* 81(6): 2817-2830.
- Bian AJ, Wang K, Kong X, Liu H, Chen F, et al. (2011) Caspase- and p38-MAPK-dependent induction of apoptosis in A549 lung cancer cells by

- Newcastle disease virus. Arch Virol 156(8): 1335-1344.
28. Krishnamurthy S, Takimoto T, Scroggs RA, and Portner A (2006) Differentially regulated interferon response determines the outcome of newcastle disease virus infection in normal and tumor cell lines. Journal of Virology 80(11): 5145-5155.
29. Zamarin D, Palese P (2012) Oncolytic Newcastle Disease Virus for cancer therapy: old challenge and new direction. Future Microbiol 7(3): 347-367.
30. Schirmacher V, Bai L, Umansky V, Yu L, Xing Y, et al. (2000) Newcastle disease virus activates macrophages for anti-tumor activity. Int J Oncol 16(2): 363-373.
31. Washburn B, Weigand MA, Grosse-Wilde A, et al. (2003) TNF-related apoptosis-inducing ligand mediates tumoricidal activity of human monocytes stimulated by Newcastle disease virus. J Immunol 170(4): 1814-1821.
32. Kaplan JM (2005) Adenovirus-Based Cancer Gene Therapy. Current Gene Therapy 5(6): 595-605.
33. Wold WS, Toth K (2013) Adenovirus vectors for gene therapy, vaccination and cancer gene therapy. Curr Gene Ther 13(6): 421-433.
34. Fenner FJ, Gibbs EPJ, Murphy FA, Rott R, Studdert MJ, et al. (2000) Veterinary Virology (2nd edn), Cademic Press, San Diego, California, USA.
35. Thacker EE, Nakayama M, Smith BF, Bird RC, Muminova Z, et al. (2009) A genetically engineered adenovirus vector targeted to CD40 mediates transduction of canine dendritic cells and promotes antigen-specific immune responses in vivo. Vaccine 27(50): 7116-7124.
36. Sorensen MR, Holst PJ, Pircher H, Christensen JP, Thomsen AR (2009) Vaccination with an adenoviral vector encoding the tumor antigen directly linked to invariant chain induces potent CD4(+) T-cell-independent CD8(+) T-cell-mediated tumor control. Eur J Immunol 39(10): 2725-2736.
37. Gabitzsch ES, Xu Y, Balint JP, Hartman ZC, Lyerly HK, et al. (2010) Anti-tumor immunotherapy despite immunity to adenovirus using a novel adenoviral vector Ad5 [E1-, E2b-]-CEA. Cancer Immunol Immunother 59(7): 1131-1135.
38. Worgall S, Busch A, Rivara M, Bonnyay D, Leopold PL, et al (2004) Modification to the capsid of the adenovirus vector that enhances dendritic cell infection and transgene-specific cellular immune responses. J Virol 78(5): 2572-2580.
39. Bartlett DL, Liu Z, Sathaiah M, Ravindranathan R, Guo Z, et al. (2013) Oncolytic viruses as therapeutic cancer vaccines. Molecular Cancer 12(1): 103.
40. Zhang L, Tang Y, Akbulut H, Zelterman D, Linton P, et al. (2003) An adenoviral vector cancer vaccine that delivers a tumor-associated antigen/CD40-ligand fusion protein to dendritic cells. PNAS 100(25): 15101-15106.
41. Perricone MA, Claussen KA, Smith KA, Kaplan JM, Piraino S, et al. (2000) Immunogene therapy for murine melanoma using recombinant adenoviral vectors expressing melanoma-associated antigens. Mol Ther 1(3): 275-284.
42. Plog MS, Guyre CA, Roberts BL, Goldberg M, St George JA, et al. (2006) Preclinical safety and biodistribution of adenovirus-based cancer vaccines after intradermal delivery. Hum Gene Ther 17(7): 705-716.
43. De Gruijl TD, van de Ven R (2012) Chapter six--Adenovirus-based immunotherapy of cancer: promises to keep. Adv Cancer Res 115: 147-220.
44. Butterfield LH, Vujanovic L (2010) New approaches to the development of adenoviral dendritic cell vaccines in melanoma. Curr Opin Investig Drugs 11(12): 1399-408.
45. Cancer Research UK (2015) Worldwide cancer statistics. London, UK.
46. CDC (Centers for Disease Control and Prevention) (2015) About Adenovirus. USA.



This work is licensed under Creative Commons Attribution 4.0 License
DOI: [10.19080/APBIJ.2018.04.555649](https://doi.org/10.19080/APBIJ.2018.04.555649)

Your next submission with Juniper Publishers will reach you the below assets

- Quality Editorial service
- Swift Peer Review
- Reprints availability
- E-prints Service
- Manuscript Podcast for convenient understanding
- Global attainment for your research
- Manuscript accessibility in different formats (Pdf, E-pub, Full Text, Audio)
- Unceasing customer service

Track the below URL for one-step submission
<https://juniperpublishers.com/online-submission.php>